



Therapeutic Potential and Utilization of Fruit Derived Waste: A Review

Bipin Bihari Panda¹, Prasanta Kumar Biswal¹, Mrs Sucheta Moharana¹

¹Gayatri College of Pharmacy, Sambalpur, Odisha, India.

ABSTRACT

Fruits have a significant role in the diet of human. Wastes are the unconsumed or unused fractions of a fruit and vegetables due to their morphological characteristics. Many fruits as well as vegetables produce up to 25- 30% of waste. The lack of proper infrastructure and methods are responsible for massive losses of important food components, by-products and residues. This review focuses on presence of bioactive compounds of some fruit by-products and possible utilization in various industries. Studies from 1986 to 2018 were reviewed from books, research paper and review articles. Currently, thousands of bioactive molecules have been isolated as well as identified from different fruit waste. The horticultural by-products are good sources of phenolic compounds, pigments, dietary fibers, organic acids, sugar derivatives, minerals and other constituents. These bioactive molecules have beneficial properties like cardioprotective, antibacterial, antimutagenic, antitumor, antiviral properties. Fruit and vegetable wastes can be utilized by isolation potential bioactive molecules and can be utilized in pharmaceutical, food, cosmetics industries. Research should be encouraged to explore the therapeutic and toxic effects, pharmacokinetics of bioactive molecules.

Key Words: Bioactive Molecules, Food-Components, Pharmacological Effects, Fruit Waste, Therapeutic Potential, Beneficial Properties

INTRODUCTION

Vegetables and Fruits have a significant role in the diet of human. Therefore, the demand of these main food commodities has increased with the growing population of world.¹ The significant amounts of fruits produced worldwide are citrus fruits, grapes, bananas, apples, mangoes, guavas, and pine-apples. Similarly, production of vegetables includes cauliflower, potato, cabbages, tomatoes, broccoli and peas.²

Food waste is part of fruits and vegetables that is not consumed and discarded due to lack of proper infrastructure and methods. The big amounts of wasted food products cause environmental problems when they decompose and emit unsafe greenhouse gases.³ Next to household garbage, fruit and vegetable processing units generate highest wastes in atmosphere.⁴

The horticultural by-products are good sources of phenolic compounds, pigments, dietary fibers, organic acids, sugar derivatives, minerals and other constituents. These compounds have beneficial properties like cardioprotective, antibacterial, antimutagenic, antitumor, antiviral properties.⁵ Fruits and vegetables like pineapples, oranges, peaches, potatoes, apples, are used for jams, juice and frozen pulp producing considerable amounts of waste materials.⁶

The Washington State Department of Ecology report exposes that decrease of waste increases profit and reduces liability. Fruit and vegetable wastes can be utilized to isolate molecules for pharmaceutical, food, cosmetics industries. So, the proper use of wastes from horticultural products may start a plan for development to reduce environmental crisis and to develop health of human being through foods having phenols, pigments, vitamins and carotenoids.⁷

Wastes are unconsumed or unused fractions of a fruit and vegetables due to their morphological characteristics. They are discarded due to lack of proper utilizations and constitute a considerable waste.⁸

Many fruits and vegetables produce up to 25- 30% of waste and are not utilized.⁹ They are rich in phenolic compounds, dietary fibers and other bioactive molecules.¹⁰ Investigations have exposed that considerable quantity of phytochemicals and necessary nutrients are available in seeds, peels those are not used as food.¹¹ Fruit peels such as lemons, grapes, oranges and jackfruit seed, longans, avocados, mangoes have above 15% more phenolic compounds than that of in pulp.¹²

Phenolic compounds are the secondary metabolites of plants responsible sensory characteristics. They add nutritional

Corresponding Author:

Prasanta Kumar Biswal, Gayatri College of Pharmacy, Sambalpur, Odisha, India; Contact detail: +91 7008767306.

Email: drprasantabiswal74@gmail.com

ISSN: 2231-2196 (Print)

ISSN: 0975-5241 (Online)

Received: 26.09.2025

Revised: 25.10.2025

Accepted: 03.11.2025

Published: 13.11.2025

quality of fruits and vegetables.¹³ Phenolic compounds are leading bioactive molecules with important biological functions.¹⁴ The compounds contain aromatic rings along with one hydroxyl groups.¹⁵ Several phenolic compounds include antioxidant property.¹⁶ The flavonoids have many subclasses like flavonols, flavones, flavanones, flavanonols, flavanols, anthocyanidins and isoflavones.¹⁷ The peel, rind, and seeds of vegetables and fruit contain high quantities of phenolic compounds.¹⁸

The wastes of fruits and vegetables contain biomolecules utilised as source of flavours and aromas. The Solid State Fermentation (SSF) is a process by which potential products are extracted.¹⁹ The market for aromas, and flavours is more due to increased demand for natural in addition to safe products. Vanillin (4-hydroxy-3-methoxybenzaldehyde) is obtained from vanillic acid. It is vital and vastly used flavour in pharmaceutical, food, cosmetic and detergent units.²⁰ Natural vanillin extraction is done from fermented pods of vanilla orchids *Vanilla planifolia*.²¹

Enzymes Amylases are of 3 types, namely α -amylase, β -amylase, and glucoamylase. Amylases are used in food processing units for products like juices, starch syrup and cakes. These are used in processes like brewing, baking, preparation of digestive aids.²² Many fruit residues are used for production of amylases.²³

Cellulases contain exo-1, 4- β -glucanase, endo-1, 4- β -d-glucanase, and β -d-glucosidase. These are utilized in food industries for extraction of aroma rich molecules and phenolic compounds from grapes pomace and others.²⁴

Similarly, invertase is employed to produce invert sugar. It has a low crystallinity in comparison to sucrose. So, it keeps the product soft and fresh for a longer period.²⁵ A peel combination in the ratio of 1 sapota: 2 pineapples: 2 bananas were used, less invertase produced in comparison to fructose as the carbon source. Invertase is exclusively used to produce jam, candies and pharmaceutical products.²⁶

Similarly, production of pectinase is done by solid-state fermentation from grape pomace using *A. awamori* yeast.²⁷ It is produced from peel of pineapple using *Penicillium chrysogenum*, pectinolytic molds and *A. Niger*.²⁸ It is produced from 6 different substrates like orange peel, lemon peel, wheat bran, banana peel, rice bran and sugarcane bagasse production using *A. niger* in solid-state fermentation.²⁹

Proteases are the principal enzymes as these are mainly used in the dairy and detergent industries.³⁰ Many agro industrial wastes also have been investigated for protease production from wastes of vegetables and fruit.³¹

Organic acids such as Citric and lactic acids are main biomolecules employed in food, cosmetic as well as pharmaceutical industries. Citric acid is produced using various molds,

yeasts as well as bacteria through fermentation. But, *A. niger* is the favourite mold for industrial production of citric acid.³²

In this review, the phytochemistry, pharmacology, traditional as well as possible industrial uses of some waste of widely used fruits are being discussed.

BANANA

Waste- Banana peel

Banana is the fruit of plant *Musa* sp. of family *Musaceae*. India is the largest producer of banana (27% of banana of world). Banana peel is main waste part of fruit. It holds nearly 30–40% (w/w) fresh. Peel composition of ripe banana is 8% protein, 6.2% ether extract, 13.8% sugars in addition to 4.8% phenolic compounds. Major components of banana peel include chlorophyll, pectin, cellulose, hemicellulose.³³

Banana pulp of *Musa Cavendish* contains 232 mg/100 g dry matter of phenolic compounds. It is about 25% of that found in peel.³⁴ It contains catecholamines, l-dopa and dopamine.³⁵ Banana waste is used as source for amylases production.³⁶ The peel contains Carotenoids like palmitate, xanthophylls, laurate. The bract contains Cyanidin and anthocyanidins like delphinine, pelargonidin, peonidin, petunidin, malvidin.³⁷

COFFEE

Waste- Coffee husk

Coffee is obtained from the cherries of plant *Coffea arabica* L. and *Coffea canephora* Pierre ex A. Froehner. The production of coffee in industry produces significant quantities of cherry husks, silver skin, cherry pulps and spent coffee.³⁸

They can be used as sorbent for the removal of heavy metals and dyes from aqueous solutions, fuel pellets production for preparing ion exchange material, spirit beverage production. They can also be used as substrate for edible mushrooms production and source of natural phenolic antioxidants, substrate for biogas, production of alcohol, biodiesel production, and biomaterial in the pharmaceutical industry.

Coffee cherry husk represents about 12 % of the berry on dry-weight basis. It contains insignificant amounts of bioactive molecules like almost 1% caffeine, 2- 5% chlorogenic acid and 5% tannins in the residue.³⁹

The high concentration of caffeine and tannins in coffee husks could be isolated for “energy drinks”. Coffee husk is used for brewed tea is called coffee cherry tea. It is useful substrate for mould, yeast and enzyme production, for its high quantity of fermentable sugars.⁴⁰

Spent coffee grounds (SCG) are the waste product from brewing coffee. These are composed of cellulose, hemicellulose,

23.9% lignin, fat, protein, and 60.46% of total dietary fibres. Spent coffee grounds is a source for biodiesel, bioethanol, steam production, spirit beverage production as source of sugars, as composting material, sorbent for metal ions removal, adsorbent for dye removal, substrate for mushroom production. It can be used as resource of phenolic antioxidant in pharmaceutical industry, food industry, polymer industry.⁴⁰ Coffee generates by-products with very rich biomolecules.⁴¹ Citric acid is cultivated from coffee husk using *A. Niger*.⁴²

PINEAPPLE

Waste- Pineapple peel, Pomace

Pineapple is the fruit obtained from plant *Ananas comosus* of family *Bromeliaceae*. India is the fourth leading producer of pineapple in world. It contributes almost 9% of total production of world of fresh pineapple.⁴³

Pineapple processing yields about 14% peels, 9% core, 15% pulp, 15% top, and 48% total final product. Pineapple peel is processed with *Penicillium chrysogenum* to get pectinase. Also acetic acid is produced from the peel by processing with *Acetobacter aceti* and *Saccharomyces cerevisiae*.⁴⁴ Pineapple peel waste has ferulic acid, which is a precursor of vanillic acid.⁴⁵ The Vanillin production from waste of pineapple is a process of 3-step.⁴⁶

Maximum amount of citric acid produced from waste of pineapple by using the yeast *Yarrowia lipolytica*, and using *A. niger* as the acting fermenter.⁴⁷

MANGO

Waste- Seed, Peel

Mango is a juicy stone fruit of *Mangifera indica* L. belongs to family *Anacardiaceae*. It has more than 1000 varieties worldwide.⁴⁸

Processing of mangoes produces about 11% peels, 13.5% seeds, 18% inoperable pulp, and 58% of finished product.⁴⁹ Hepatoprotective activities in mango seed kernels were studied by Nithitanakool et al. in 2009.⁵⁰ The anti-diarrheal activity of extracts of seeds of *M. indica* was investigated.⁵¹ Mango kernel aqueous extract has anti-diarrheal activity at 0.25 - 0.50 mg/ml dose.⁵²

Mango peel has 51.2% dry matter of Total Dietary Fibers. It contains 32% dry matter insoluble fibers and 19% dry matter soluble fibers.⁵³ The dietary fiber of mango peels contains Total Dietary Fibers content 40.6- 72.5%, with glucose, galactose, and arabinose as chief neutral sugars in insoluble and soluble dietary fibers.⁵⁴ "Tommy Atkins" variety mango has 28.05% dry matter of dietary fibers, in which 13.80%

dry matter insoluble and 14.25% dry matter soluble fibers.⁵⁵

The mango kernel contains 2% crude fiber.⁵⁶ Ashoush et al. found nearly 0.26g/100 g crude fiber in mango kernel powder and nearly 9.33 g/100 g crude fiber in mango peel powder. Mango kernels are used as resource for amylases production.⁵⁷

GRAPE

Waste- Seed, Pomace

Vitis vinifera is a perennial, woody climbing vine. Its processing industries produce about 5 to 9 MMT of solid waste every year worldwide. It constitutes 20- 30% of processed materials.⁵⁸ Grapes pomace is a rich source of dietary fibers, like hemicelluloses, cellulose, and small amount of pectins.⁵⁹

Grape seeds as well as skins are by-products of wine and juice industries. They contain monomeric, oligomeric, as well as polymeric proanthocyanidins.⁶⁰

Grape seeds contain flavonoids (4- 5%), including kaempferol-3-O-glucosides, quercetin- 3-O-glucosides, quercetin and myricetin. 60-70% of polyphenols are found in seeds of grape. The grape seed polyphenols are flavan-3-ol derivatives. The major compounds are (+)-catechins, (dimers (B1-B5), procyanidins C1, and procyanidin B5- 32 -gallate. Grape seeds contain procyanidins or proanthocyanidins.⁶¹

Grape seeds contain 6-20% oil utilized as edible oil, soaps, linseed substitute. *Amalagma* made from the seed is a folk remedy for condylomata of the joints. Grape seed extract has antioxidant as well as free radical scavenging property.⁶²

The sparing or recycling effect of procyanidins from *V. Vinifera* seeds on alpha-tocopherol was established in phosphatidylcholine liposomes and red blood cells. Other than free radicals scavenging nature, Procyanidins also inhibit xanthine oxidase activity. Xanthine oxidase enzyme triggers the oxy-radical cascade.⁶³

APPLE

Waste- Peel, Pomace

Apple is the fruit obtained from plant *Malus domestica* belongs to family *Rosaceae*. It is cultivated commercially in 91 countries.⁶⁴

Apples generate 89.09% final products and 10.91% seed and pulp as waste during slicing. Apple peel has 0.91% FW more dietary fiber content than pulp. The insoluble and soluble dietary fibers are 0.46% FW and 0.43% FW respectively.⁶⁵ Apple peels contain up to 3300 mg/100 g dry matter of phenolic content.⁶⁶ The pineapple flavour constituent is ethyl butyrate.

Apple pomace is used as substrate with *A. niger* to generate citric acid.⁶⁷

The apple peels contain compounds such as catechin, procyanidins, epicatechin. It also contains chlorogenic acid, quercetin conjugates, phloridzin. Though apple flesh contains catechin, epicatechin, procyanidin and phloridzin, these molecules are present in lower concentrations than in peels. Only Chlorogenic acid found is higher in flesh than in peel.⁶⁸

Apple peels contain higher antioxidant compound, quercetin. The peel has more antioxidant activity and more bioavailability than the apple flesh. Whole apples with peels are better capable for inhibition of cancer cell proliferation compared to apples without peels. Apple peels contain two to six times more phenolic compounds than in flesh. 2-3 times more flavonoids present in peels as compared to flesh depending on variety.⁶⁹

BER

Waste- Seed

Ber fruit is obtained from the plant *Ziziphus mauritiana* of family *Rhamnaceae*. The seed is sedative, hypnotic, narcotic, stomachache as well as tonic. It is administered internally in the treatment of insomnia, palpitations, nervous exhaustion, night sweats in addition to excessive perspiration.⁷⁰

There are 2 types of domestic jujubes, i.e. *Z. mauritiana* Lam. the Indian jujube (Ber), and *Z. jujuba* Mill, (common jujube).

Flavonoid glycosides spinosins was extracted from seeds of *Z. jujuba* var. Three types of acylated flavone-C glycosides were isolated. These molecules have sedative activity in rats. The saponins found from of *Z. jujuba* seeds are Jujubosides A and Jujubosides B.⁷² Sedative flavonoids like Swertisin as well as spinosin were extracted from fruit and seeds of *Z. jujuba*.

The leaves and seeds of several *Ziziphus* species have anxiolytic and hypnotic-sedative property. They depress the activity of CNS and reduce anxiety, induce sleep. It concludes to produce sleep, but not muscle relaxant or anticonvulsant.⁷¹ The inhibitory potential of Jujuboside A on rat hippocampus is revealed.⁷² Anxiolytics effect of a polyherbal formulation having seed extract of *Z. jujuba* is reported in mice.⁷³

JACKFRUIT

Waste- peel

Jack fruit is obtained from the plant *Artocarpus heterophyllus* belongs to family *Moraceae*. It is native to India.⁷⁴ In addition to India, the fruit is commonly cultivated in Sri Lanka, Bangladesh, Burma, Indonesia, Philippines, Thailand, Brazil,

Malaysia.⁷⁵ In India, it is distributed in many states.⁷⁶ It is considered as the Poor man food.⁷⁷

Polyphenol as well as flavonoid are present in spine, skin and rind, respectively. All the three extracts have anti-inflammatory properties comparable to diclofenac in both *in-vitro* and *in-vivo* models. The skin revealed maximum anti-inflammatory property. The rind of fruit had favoured cyclooxygenase-2 inhibition. The spine as well as skin reduced both cox-1 as well as cox-2 *in-vitro*.⁷⁸ Jackfruit peel can be employed as adsorbent for removal of cadmium.⁷⁹

JAMUN

Waste- Seed

Jamun is obtained from the plant *Syzygium cumini* of family *Myrtaceae*. It is termed as Jambolan, amblang, Kala Jamun, Jambul in India. It is advised as a safe drug in different diseases by health organizations.⁸⁰ This plant is also used for treatment, prevention of many diseases in Homoeopathy practice in different countries.

The seed of fruit is used as alternative natural healing medicine in Unani, Chinese and Ayurvedic system of medicines. The bark and seeds are famous in Far East for treatment of dysentery. They are used in the control of hyperglycaemia, glycosuria in patients of diabetes.

Syzygium cumini seeds are used for various disorders as anti-inflammatory, hypolipidaemic, antidiabetic, antioxidant, neuroprotective. Seeds proved to have the DNA protection effect against radiation.⁸¹ The cardioprotective property of seeds of *Syzygium cumini* (Jamun) revealed against the DOX induced acute myocardial stress in rats.⁸²

Ethanol extract of seed kernel of *Syzygium cumini* reduces thiobarbituric acid reactive substance (TBARS), glutathione (GSH), superoxide dismutase (SOD) and catalase (CAT) activity.

The seeds contain crude fiber (16.9%), Protein (6.3 to 8.5%), Fat (1.18%). It contains Magnesium 17 mg, Phosphorus (2%), Potassium 79 mg (2%), Sodium 14 mg (1%). More anti-inflammatory activities are in stem and seeds of *Syzygium cumini*.⁸³ The chloroform extracts of dried seeds showed antipyretic activity. The seed extract is active versus yeast induced pyrexia.³³

The *Syzygium cumini* seed extracts contain alkaloids, glycosides, steroids, triterpenoids, saponins, tannins, flavonoids and carbohydrates.⁸⁴

LITCHI

Waste- Peel, Seed

Litchi fruit is obtained from the plant *Litchi chinesis* Sonn of the family *Sapindaceae*. The peel of fruit is tough, inedible,

covered with sharp protuberances and possesses a profound pink-red colour when ripe. Processing and consumption of the fruit generates by-products such as epicarps and seeds, which are often discarded.

The seeds may be used as a food additive in pharmaceutical drugs. Studies on plant extracts and fruit based wastes as antioxidants for preserving foods as well as neutralizing free radicals in the human body are being developed.⁸⁵

Peel and seed of litchi possess high energy and nutritional value. They are loaded in antioxidants like ascorbic acid and phenolic compounds particularly gallic acid, flavonoids like procyanidin B4, procyanidin B2, epicatechin cyanidin-3-retinoside, cyanidin-3-glucoside, quercetin-3-retinoside and quercetin-3-glucoside.⁸⁶ They also contain anthocyanins.⁸⁷

By-products of litchi possess antioxidant, anti-inflammatory, anti-hyperlipidemia, anti-hyperglycaemic, hepatic and cardioprotective properties.⁸⁸ The ethanomedicinal use of *L. Chinensis* kernel oil on cardiovascular disorder may be because of its capability of reducing total cholesterol, LDL and increasing HDL values.⁸⁹ Aqueous extract isolated from seed increases insulin sensitivity and decreases blood glucose levels, triglyceride, leptin, tumor-necrosis factor in Type-2 diabetes mellitus in rat model.⁹⁰

TAMARIND

Waste- Seed

The fruit Tamarind is obtained from the plant *Tamarindus indica*. It is a famous as well as common tree found in India.⁹¹ It is used as a stabilizer, flavour and binder in preparation of food. Generally, people use Imli to prepare pickles, curries, jam and sauce.

The seeds are used to make flour for making bread in famine seasons. The seeds are fried and consumed by poorer.⁹² It is very useful for people of hot area and near to equator.⁹⁴

Tamarind seed powder was used for tannase production by *A. niger*, and the seeds act as anti-asthmatic, antiulcer as well as antioxidant agent.⁹³ Vaginal Atony is treated by a pessary of seed kernel. The fried seed paste was used on anus after setting the tract in proper position for treatment of rectal prolapse. In Unani system, spermatorrhoea, nocturnal emissions as well as seminal debility are cured by use of roasted kernel and seeds. Homeopathic system uses seed for the treatment of stomachache.⁹⁴ It is used in treatment of cough and for uvula relaxation.⁹⁵ The urethral discharge and polyuria are treated by administration of seed kernel with milk.⁹⁶ The red outer covering of seeds is very useful in diarrhoea and dysentery.⁹² The seeds are used for treatment of colitis and other disorders of intestine.⁹⁷ Seeds are helpful in vaginal discharges as well as ulcers.⁹⁸

Seeds are rich in phenolic compounds, polymeric tannins, glycosides, fatty acids, flavonoids, saponins, alkaloids.⁹⁴ There is also presence of fatty acids, essential amino acids in seeds.⁹⁹ Tamarind seeds contain 2-hydroxy-3', 4'-dihydroxyacetophenone (TAO), 3, 4-dihydroxyphenylacetate (TA2) and (-)-epicatechin. Seed consist of acetic acid, arabinose, dihydroxyphenyl acetate.¹⁰⁰ Oil obtained from seeds consists of lauric acid, myristic acid, palmitic acid, stearic acid, arachidic acid, lignoceric acids in addition to linoleic acids. roasted kernel includes calcium 121 mg and phosphorous 237 mg per 100 gm. It also contains proteins like prolamines and albumins.⁹⁵

Methanolic extract of seeds of *Tamarindus indica* showed analgesic as well as anti-inflammatory effects in inhibiting carrageenan induced paw edema in rats.⁹⁴ The seed aqueous extract of *Tamarindus indica* proved to have potent anti-diabetogenic activity which reduces blood sugar concentration in streptozotocin induced diabetic male rat. Aqueous extract 80 mg/100 g body weight / day in streptozotocin induced diabetic rat showed reduction of fasting blood sugar concentration after 7 days of continuous treatment.¹⁰⁰ Anti-inflammatory activity is supported by presence of saponins, flavonoids and tannins in seeds of *Tamarindus*.⁹⁴

The ethanolic extract of *Tamarindus indica* seed coat revealed potent antioxidant property. Antiulcer property discovered in methanolic extract of seed coats of Tamarind. The result revealed diminution in total gastric juice volume and total acidity in ulcer by pylorus ligation method. Also, there was significant decrease in ulcer index.¹⁰¹ Aqueous extract of various parts of *Tamarindus indica* like fruits, leaves as well as unroasted seeds revealed significant hepatoprotective activity in Paracetamol induced hepatotoxicity in rats.¹⁰²

ORANGE

Waste- Peel

Orange is the fruit obtained from the tree *Citrus sinensis*. The production of orange is about 70% of the total production of Citrus fruit.¹⁰³ It is indigenous to Asia. Now it is widespread all over the world.¹⁰⁴

The orange peel contains 57% Dry Weight Total Dietary Fibres, of which 9.41% is soluble fraction and 47.6% insoluble fraction. The chief components of fibers are cellulose as well as pectin polysaccharides.¹⁰⁵

L-Rhamnose is the chief component of cell wall pectins. It is used for the generation of the strawberry flavour "furanol" as raw material.¹⁰⁶

Orange peel gives the best result for pectinase production. The Enzyme Pectin has many important uses in fruit juices industries as well as wines industries for extraction, clarification

and concentration. Also, extraction of flavours, pigments, and essential oils is achieved by these enzymes from the residues of plant.¹⁰⁷

Silver nanoparticles were synthesized using aqueous extract of peel of *C. Sinensis*. It demonstrated zones of inhibition against *Escherichia coli*, *Pseudomonas aeruginosa* and *Staphylococcus aureus*.¹⁰⁸

Hexane extract of Peel of *C. sinensis* revealed antimycobacterial activity against drug-sensitive, Isoniazid-resistant, and ethambutol-resistant variants of *M. tuberculosis*.¹⁰⁹

Polymethoxylated flavones obtained from *Citrus sinensis* peel extract such as flavone-7-O-[6-acyl]-glucoside, natsudaic acid, tetramethyl-O-scutellare in, nobletin, tangeretin, heptamethoxy flavone revealed activity against *Aspergillus Niger* using a microbroth dilution assay.¹¹⁰

Essential oil obtained from peels of *Citrus sinensis* obtained by cold-pressing method revealed activity against *Mucor hiemalis*, *P. expansum* as well as *F. proliferatum*.¹¹⁴

Polymethoxy flavones obtained from *C. sinensis* peels showed activity on lung cancer cells of human. The four flavonoids showed similar action in lung cancer cells H441 in addition to H460 of human.¹¹⁰

Orange peel oil obtained by Cold-pressed method contains mixture of 75.1% non-hydroxylated Polymethoxy flavones in addition to 5.44% hydroxylated Polymethoxyflavones. A mixture of 97.2% hydroxylated Polymethoxyflavones trigger apoptosis of breast cancer cells MCF-7.¹¹¹ 5% of orange peel extract containing 30% Polymethoxyflavones treated to mice it decreases the tumors development.¹¹²

Flavonoid content of some fractions of methanolic extract of peel of Navel orange has antioxidant property *in vitro*. The major flavonoids were O-glycosylated flavones, Polymethoxylated flavones, O-glycosylated flavanones, C-glycosylated flavones, O-glycosylated flavanols in addition to phenolic acids with their ester derivatives. Hesperidin is chief flavonoid glycoside of peel of orange. Antioxidant property of fractions of methanolic extract of orange peel was estimated. It was proved that the fractions have moderate antioxidant activity in comparison to the activity given by the aglycones, diosmetin and hesperetin. Aglycones revealed a significant more hydroxyl scavenging activity as compared to quercetin.¹¹³

LEMON

Waste- Peel

Lemon is the fruit of *Citrus limon* which is a small evergreen tree originated from Asia. This flowering plant belongs to family *Rutaceae*. Nearly 1300 species are available in genus

citrus.¹¹⁴

In both the leaves and peel of the *citrus limon*, essential oils, limonene is the chief element. Peel oil contains μ -terpinene, β -pinene, myrcene.¹¹⁵

Nearly 8.452 % w/w Ca is present in lemon peel which helps in bone formation, controls several cell procedures and plays important structural functions in living organisms.¹¹⁶

Lemon peel eating reduces cholesterol effectively.¹¹⁷ The research reveals that lemon peel is also a useful antimicrobial agent and an astringent.¹¹⁸

The dietary fiber amount is more in the pulp (77.93%) than peels (53.02%). The soluble fraction is 27.91% in pulp and 18.48% in peels. Soluble fraction in range of 34.5% in the peel and 35.8% in pulp of the TDF is best for human consumption.¹¹⁹ Citrus waste is used as substrate for amylase production.¹²⁰

PAPAYA

Waste- Peel, Seed

Papaya is the fruit obtained fruit *Carica papaya* Linn belongs to family *Caricaceae*. It is recommended for consumption for prevention of vitamin A deficiency. The unripe fruit is good source of vitamin A.¹²¹

The fruit produces about 8.5% peel, 6.5% seeds, 32% unusable pulp and nearly 53% final product during dicing. Flavonoid, Sterol is available in unripe papaya peel extract however not in unripe papaya seed extract. Steroids, saponins, sterols and flavonoid are present in unripe papaya peel extracts. Saponins, tannins, steroids and terpenoids seemed to be positive in unripe papaya seed extracts. Extract of papaya seed (unripe) is richer in total phenol content in comparison to papaya peel. Also the unripe papaya seed is rich in total flavonoid in comparison to unripe papaya peel extract. But the results showed that unripe papaya peel extract has more ferric reducing antioxidant nature in comparison to extract of unripe papaya seed. The results revealed that more total phenolic compound in unripe papaya seed as compared to unripe papaya peel. So extract of unripe papaya seed maybe excellent sources of dietary antioxidant.¹²²

DATE

Waste- Seed

Date is obtained from the plant *Phoenix dactylifera* belongs to family *Arecaceae*. Seeds of Date are good source of antioxidants as well as phenolic compounds.¹²³ Seed oil has more phenolic content in comparison to other edible oils except olive oil.¹²⁴ Waste obtained from Date is used as substrate for amylases production.¹²⁵

Date seeds are proved to contain higher amounts of flavonoids, alkaloids, anthraquinone, saponin, tannin and terpenoids. The mineral contents of seeds of date are significant. Date seeds have high potassium content than sodium content.¹²⁶

The higher potassium to sodium ratio can give health benefit in patients with cardiovascular disorder who take extract of date palm seed as coffee. The antioxidant activity of date seeds is less than leaf extracts of Algerian date palm.¹²⁷ The cerebroprotective effects of date seed extract were examined in rats affected by cerebral ischemia. Seed extract decreases neuronal damage. Ultra structures of cortical neurons were protected by seed extract. This cerebroprotective capability of Date seed is due to its antioxidant activity. Date seed extract attenuates muscle weakness which proved protective efficiency against damage due to ischemic reperfusion.¹²⁸

POMEGRANATE

Waste- Peel

Pomegranate fruit is obtained from the plant *Punica granatum* belongs to family *Lythraceae*. It is indigenous to Iran and it extended to India. It is commonly known as Anaar in native language.¹²⁹

Pomegranate seed oil contains Punicic acid, ellagic acid, fatty acids and sterols. Pomegranate pericarp such as rind, peel contain Phenolic molecules such as punicalagins, catechin, quercetin, rutin, anthocyanidins, gallic acid, other flavonoids.¹³⁰

The pomegranate seeds contain poly unsaturated fatty acids such as linoleic acid, palmitic acid, oleic acid, other lipids punicic acids, stearic acid and linolenic acid.¹³¹

Peel of pomegranate is about half of total fruit weight which is contain bioactive compounds, potassium, nitrogen, magnesium, calcium like minerals, phosphorus, sodium, proanthocyanidins, flavonoids, ellagitannins and phenolic molecules.¹³² It also contains polysaccharide molecules.¹³³ The seeds contain sex steroid estrone, phytoestrogen coumestrol, pectin, minerals, crude fibers, proteins, vitamins, sugars. Seeds have isoflavones mostly polyphenols and genistein.¹³⁴

Pomegranate peels extract triggers synthesis of type I procollagen as well as inhibits matrix metalloproteinase-1 enzyme and interstitial collagenase synthesis by dermal fibroblasts. Seed oil of pomegranate has stimulatory effect on proliferation of keratinocytes in monolayer culture. It indicates pomegranate peel aqueous extract increases the regeneration of dermis and seed oil of pomegranate improves the regeneration of epidermis.¹³⁵

The antidiarrheal efficacy of peel extract of pomegranate was assessed in rats by administration of 400 mg/kg through oral

route and concluded that it decreases number of defecations and weight of feces.¹³⁶ Another study revealed antidiarrheal activity of aqueous extract of peels of pomegranate in rats.¹³⁷

DISCUSSION

The advantages of the bioactive compounds in these wastes are main reason for significant number of research studies. This review emphasizes the nature, types of waste obtained from fruits. The review discusses the bioactive molecules such as phenolic compounds, dietary fibers, vitamins, flavours, enzymes, minerals available in fruit waste.

Citrus peel contains more quantity of polyphenols as compared to the edible part of fruit.¹⁵ It indicates that they could be inexpensive and readily available resources of bioactive compounds for use in the food and pharmaceutical industries.

The cerebroprotective effects of date seed extract were examined in rats affected by cerebral ischemia and found to decreases neuronal damage. Similarly, anxiolytic effect of a polyhedral formulation having seed extract of *Z. jujuba* is reported in mice.⁷⁵ In a study, the seed aqueous extract of *Tamarindus indica* proved to have potent antidiabetogenic activity.¹⁰⁰

Extracted compounds can be used in food, pharmaceuticals, cosmetic, and chemical industries. They can also be used in food research in development of functional foods.

CONCLUSIONS

The researches in the field of by-products of fruit and vegetable have developed in past few years. Regarding pharmaceutical applications, numerous reviews related to bioactive compounds have been published. Understanding the benefits of polyphenols is difficult study. But it is essential for dietary recommendations. Numerous evidences have been generated by *in vitro* studies, animal studies in addition to epidemiological approaches. But the concrete proof from clinical trials is very limited. More data from well-designed long-term clinical trials are essential to confirm the results obtained from *in vitro* studies and animal studies.

It can be concluded that, by-products of fruit are important source of bioactive compounds. These could be better utilised instead of ending up as wastes. Besides utilizing them as animal feed or for methane production through anaerobic fermentation, these can be used in food, cosmetic as well as pharmaceutical industries. Interdisciplinary approaches must be carried out, correlated with efficient public awareness campaigns to reduce food waste. We will be able to prescribe the proper dietary recommendations in favour of a particular

health benefit only after establishment of most effective concentration of a specific compound.

ACKNOWLEDGEMENT

Authors acknowledge the immense help received from the scholars whose articles are cited and included in references of this manuscript. The authors are also grateful to authors and editors of all those articles, journals and books from where the literature for this article has been reviewed and discussed.

Source of funding:

This research received no external funding.

Conflict of Interest:

The authors deny any conflict of interest

Authors' Contribution:

Conceptualization: Bipin Bihari Panda

Investigation: Prasanta Kumar Biswal

Supervision: Sucheta Moharana

REFERENCES

- Schieber A, Stintzing FC, Carle R. By-products of plant food processing as a source of functional compounds - recent developments. *Trends Food Sci Technol*. 2001;12:401–13.
- Socaci SA, Rugina DO. Antioxidant Compounds Recovered from Food Wastes. *IntechOpen*. 2017; chapter1: 1-21.
- Venkat K. The climate change and economic impacts of food waste in the United States. *Intl J Food Sys Dyn*. 2011; 2:431–46.
- Gowe C. Review on potential use of fruit and vegetables by-products as a valuable source of natural food additives. *Food Sci Qual Manag*. 2015; 45: 47–61.
- Dilas S, Anadanovic-Brunet J, Etkovic G. By-products of fruits processing as a source of phytochemicals. *Chem Ind Chem Engr Quart*. 2009; 15:191–202.
- Rodriguez R, Jimenez A, Guillen R, Heredia A. Postharvest changes in white asparagus cell wall during refrigerated storage. *J Agric Food Chem*. 1999; 47:3551–7.
- WSDE. Pollution prevention in fruit and vegetable food processing industries. Publication No. 94-056; 1994. Olympia, WA: Washington State Department of Ecology. Available from: <https://apps.ecology.wa.gov/publications/SummaryPages/94056.html>.
- Panouille M, Ralet MC, Bonnin E, Thibault JF. Recovery and reuse of trimmings and pulps from fruit and vegetable processing. In: Waldron KW, editor. *Handbook of waste management and co-product recovery in food processing*. Cambridge: Woodhead Publishing Limited. 2007;417–47.
- Ajila CM, Aalami M, Leclavathi K, Rao UP. Mango peel powder: a potential source of antioxidant and dietary fiber in macaroni preparations. *Innov Food Sci Emerg Technol*. 2010;11:219–24.
- Galanakis CM. Recovery of high added-value components from food wastes: conventional, emerging technologies and commercialized applications. *Trends Food Sci Technol*. 2012; 26:68–87.
- Rudra SG, Nishad J, Jakhar N, Kaur C. Food industry waste: mine of nutraceuticals. *Intl J Sci Environ Technol*. 2015;4:205–29.
- Soong YY, Barlow PJ. Antioxidant activity and phenolic content of selected fruit seeds. *Food Chem*. 2004; 88:411–7.
- Lapornik B, Prosek M, Wondra AG. Comparison of extracts prepared from plant by-products using different solvents and extraction time. *J Food Engr*. 2005; 71:214–22.
- Ignat I, Volf I, Popa VI. A critical review of methods for characterisation of polyphenolic compounds in fruits and vegetables. *Food Chem*. 2011; 126:1821–35.
- Balasundram N, Sundram K, Samman S. Phenolic compounds in plants and agri-industrial by-products: antioxidant activity, occurrence, and potential uses. *Food Chem*. 2006; 99:191–203.
- Heim KE, Tagliaferro AR, Bobilya DJ. Flavonoid antioxidants: chemistry, metabolism and structure-activity relationships. *J Nutr Biochem*. 2002; 13:572–84.
- Robbins RJ. Phenolic acids in foods: an overview of analytical methodology. *J Agric Food Chem*. 2003; 51:2866–87.
- Friedman M. Chemistry, biochemistry, and dietary role of potato polyphenols: a review. *J Agric Food Chem*. 1997; 45:1523–40.
- Zheng Z, Shetty K. Solid-state production of beneficial fungi on apple processing wastes using glucosamine as the indicator of growth. *J Agric Food Chem*. 1998; 46:783–7.
- Tilay A, Bule M, Kishenkumar J, Annappure U. Preparation of ferulic acid from agricultural wastes: its improved extraction and purification. *J Agric Food Chem*. 2008; 56:7644–8.
- Panouille M, Ralet MC, Bonnin E, Thibault JF. Recovery and reuse of trimmings and pulps from fruit and vegetable processing. In: Waldron KW, editor. *Handbook of waste management and co-product recovery in food processing*. Cambridge: Woodhead Publishing Limited. 2007; 417–47.
- Khan G J, Jamshaid M, Sajid MI. The Pharmacological, Physiological and Toxicological Effects of Pomegranate Fruit Extract and Its Constituents. *Canadian J Appl Sci*. 2014;3 (4): 66-80
- Cyprian EO, Henrietta OO, Omonigho SE. Fruit Wastes as substrate for the production of Amylase by *Aspergillus niger*. *Trop J Nat Prod Res*, 2017; 1(4):182-185
- Meyer AS, Jepsen SM, Sorensen NS. Enzymatic release of antioxidants for human low-density lipoprotein from grape pomace. *J Agric Food Chem*. 1998; 46:2439–46.
- Sagar NA, Pareek S, Sharma S, Yahia E M., Maria GL. Fruit and Vegetable Waste: Bioactive Compounds, Their Extraction, and Possible Utilization. *Compr Rev Food Sci Food Saf*. 2018;17: 512.
- Panda SK, Mishra SS, Kayitesi E, Ray RC. Microbial-processing of fruit and vegetable wastes for production of vital enzymes and organic acids: biotechnology and scopes. *Environ Res*. 2016; 146:161–72.
- Botella C, De Ory I, Webb C, Cantero D, Blandino A. Hydrolytic enzyme production by *Aspergillus Awamori* on grape pomace. *Biochem Engr J*. 2005; 26:100–6.
- Okafor UA, Okochi VI, Chinedu SN, Ebuechi OAT, Pectinolytic activity of wild-type filamentous fungi fermented on agro-wastes. *Afr J Microbiol Res*. 2010; 4:2729–34.
- Mrudula S, Anitharaj R. Pectinase production in solid state fermentation by *Aspergillus niger* using orange peel as substrate. *Glob J BiotechnolBiochem*. 2011; 6:64–71.
- Kirk O, Borchert TV, Fuglsang CC. Industrial enzyme applications. *Curr Opin Biotechnol*. 2002; 13:345–51.
- Bharathiraja S, Suriya J, Krishnan M. Production of enzymes from agricultural wastes and their potential industrial applica-

- tions. Adv Food Nutr Res. 2017;80:125-48.
32. Ali BH, Bashir AK, Tanira MOM. The effect of *Rhazya stricta* Decne, a traditional medicinal plant, on the forced swimming test in rats. Pharmacol Biochem Behav. 1998;59(2):547-50.
 33. Mahapatra PK, Chakraborty D, Chaudhari AKN. Anti-inflammatory and antipyretic activities of *Syzygium cumini*. Planta Med. 1986;52(6):540.
 34. Someya S, Yoshiki Y, Okubo K. Antioxidant compounds from bananas (*Musa cavendish*). Food Chem. 2002; 79:351-4.
 35. Gonzalez-Montelongo R, Lobo MG. Antioxidant activity in banana peel extracts: testing extraction conditions and related bioactive compounds. Food Chem. 2010;119:1030-9.
 36. Unakal C, Kallur RI, Kaliwal BB. Production of alpha-amylase using banana waste by *Bacillus subtilis* under solid-state fermentation. Eur J Exp Biol. 2012; 2:1044-52.
 37. Reddy GV, Babu PR, Komaraiah P, Roy KRRM, Kothari IL. Utilization of banana waste for the production of lignolytic and cellulolytic enzymes by solid substrate fermentation using two *Pleurotus* species (*P. ostreatus* and *P. sajor-caju*). Process Biochem. 2003;38:1457-62.
 38. Matsuda H, Matsuda T, Ikebata A, Yamahara J. Bioactive saponins and glycosides. XIV. Chem Pharm Bull. 1999;47:12-4.
 39. Franca AS, Oliveira LS. Coffee processing solid wastes: current uses and future perspectives. In: Ashworth GS, Azevedo P, eds. Agricultural wastes. New York: Nova Science; 2009. p. [chapter 8 pages].
 40. Lenka B, Maros S, Alica B. Review: utilization of waste from coffee production. Fac Mater Sci Technol. 2017;25(40):91-101.
 41. Ali A, Naheed B, Muhammad T. Phytochemical and therapeutic evaluation of date (*Phoenix dactylifera*): a review. J Pharm Altern Med. 2016; 9:11-7.
 42. Prabha MS, Rangaiah GS. Citric acid production using *Ananas comosus* and its waste with the effect of alcohols. Int J Curr Microbiol App Sci. 2014;3:747-54.
 43. Chaudhary V. Pineapple (*Ananas comosus*) product processing: a review. J Pharmacogn Phytochem. 2019;8(3):4642-52.
 44. Woo WS, Kang SS, Shim SH. The structure of spinosin (2''-O-beta-glucosylswertisin) from *Ziziphus vulgaris* var. *spinosus* seeds. Phytochemistry. 1979;18(2):353-5.
 45. Priefert H, Rabenhorst J, Steinbuechel A. Biotechnology production of vanillin. Appl Microbiol Biotechnol. 2001;56:296-314.
 46. Lun OK, Wai TB, Ling LS. Pineapple cannery waste as a potential substrate for microbial biotransformation to produce vanillic acid and vanillin. Int Food Res J. 2014;21:953-8.
 47. Imandi SB, Bandaru VVR, Somalanka SR, Bandaru SR. Application of statistical experimental designs for the optimization of medium constituents for the production of citric acid from pineapple waste. Bioresour Technol. 2008; 99:4445-50.
 48. Solis-Fuentes JA, Duran-de-Bazua MC. Mango (*Mangifera indica* L.) Seed and its fats. In V. Preedy, R. R. Watson, & V. B. Patel (Eds.), Nuts and Seeds in health and disease prevention Chapter 88. San Diego: Academic Press 2011; 741-748.
 49. Joshi VK, Kumar A, Kumar V. Antimicrobial, antioxidant and phytochemicals from fruit and vegetable wastes: a review. Int Food Ferment Technol. 2012;2:123-36.
 50. Nithitanakool S, Pithayanukul P, Bavovada R. Antioxidant and hepatoprotective activities of Thai mango seed kernel extract. Planta Med. 2009;75:1118-23.
 51. Sairam K, Hemalatha S, Kumar A, Srinivasan T, Ganesh J. Evaluation of anti-diarrheal activity in seed extracts of *Mangifera indica*. J Ethnopharmacol. 2003;84:11-5.
 52. Alkizim FO, Matheka D, Abdulrahman FK, Muriithi A. Inhibitory effect of *Mangifera indica* on gastrointestinal motility. Med Chem Drug Discov. 2012;2(1):9-16.
 53. Ajila CM, Leelavathi K, Rao UP. Improvement of dietary fiber content and antioxidant properties in soft dough biscuits with the incorporation of mango peel powder. J Cereal Sci. 2008;48:319-26.
 54. Ajila CM, Prasada Rao UJS. Mango peel dietary fibre: composition and associated bound phenolics. J Funct Foods. 2013;5:444-50.
 55. Vergara-Valencia N, Granados-Perez E, Tovar J, Ruales J. Fibre concentrate from mango fruit: characterization, associated antioxidant capacity and application as a bakery product ingredient. LWT Food Sci Technol. 2007;40:722-9.
 56. Nzikou JM, Kimbonguila A, Matos L. Extraction and characteristics of seed kernel oil from mango (*Mangifera indica*). Res J Environ Earth Sci. 2010;2:31-5.
 57. Rudra SG, Nishad J, Jakhar N, Kaur C. Food industry waste: mine of nutraceuticals. Int J Sci Environ Technol. 2015;4:205-29.
 58. Kirtikar KR, Basu BD. Indian medicinal plants. Dehradun: International Book Distributors; 1991. p. 887-91.
 59. Kammerer D, Claus A, Schieber A. A novel process for the recovery of polyphenols from grape (*Vitis vinifera* L.) pomace. J Food Sci. 2005;70:C157-63.
 60. Torres JL, Bobet R. New flavanol derivatives from grape (*Vitis vinifera*) by-products. Antioxidant aminoethylthio-flavan-3-ol conjugates from a polymeric waste fraction used as a source of flavanols. J Agric Food Chem. 2001;49:4627-34.
 61. Marjan N, Hossein H. Review of the pharmacological effects of *Vitis vinifera* (grape) and its bioactive compounds. Phytoter Res. 2009;23:1197-204.
 62. Jayaprakasha GK, Selvi T, Sakariah KK. Antibacterial and antioxidant activities of grape (*Vitis vinifera*) seed extracts. Food Res Int. 2003;36:117-22.
 63. Facino RM, Carini M, Aldini G. Free radicals scavenging action and antienzyme activities of procyanidines from *Vitis vinifera*: a mechanism for their capillary protective action. Arzneimittelforschung. 1994;44:592-601.
 64. Gorinstein S, Zachwieja Z, Foltá M, Barton H. Comparative contents of dietary fiber, total phenolics, and minerals in persimmons and apples. J Agric Food Chem. 2001;49:952-7.
 65. Yoshikawa M, Murakami T, Ikebata A. Bioactive saponins and glycosides. X. On the constituents of *Ziziphus spinosus* seeds, the seeds of *Ziziphus jujuba* Mill. var. *spinosa* Hu (1): structures and histamine release-inhibitory effect of jujubosides A1 and C and acetyljujuboside B. Chem Pharm Bull. 1997;45:1186-92.
 66. Wolfe KL, Liu RH. Apple peels as a value-added food ingredient. J Agric Food Chem. 2003;51:1676-83.
 67. Dhillon GS, Brar SK, Verma M. Enhanced solid-state citric acid bio-production using apple pomace waste through surface response methodology. J Appl Microbiol. 2011;110:1045-55.
 68. Lee K, Kim Y, Kim D, Lee H, Lee C. Major phenolics in apple and their contribution to the total antioxidant capacity. J Agric Food Chem. 2003; 51:6516-20.
 69. Eberhardt M, Lee C, Liu RH. Antioxidant activity of fresh apples. Nature. 2000;405:903-4.
 70. Palejkar CJ, Palejkar JH, Patel AJ. A plant review on *Ziziphus mauritiana*. Int J Univ Pharm Life Sci. 2012;2(2):202.
 71. Woo WS, Kang SS, Shim SH. The structure of spinosin (2''-O-beta-glucosylswertisin) from *Ziziphus vulgaris* var. *spinosus* seeds. Phytochemistry. 1979;18(2):353-5.
 72. Zeng L, Zhang RY, Wang X. Studies on the constituents of *Ziziphus spinosus* Hu. Acta Pharm Sin. 1987; 22:114-20.
 73. Huang PW, Lin YC. Anxiolytic effect of seed of *Ziziphus jujuba* in mouse models of anxiety. J Ethnopharmacol. 2000;72(3):435-41.

74. Feng ZY, Zheng XX. The effect of Jujuboside A on the evoked field potentials of granule cells in dentate gyrus. *J Zhejiang Univ Sci.* 2002;3:9-12.
75. Lin YC, Peng YS. Anxiolytic effect of seed of *Ziziphus jujuba* in mouse models of anxiety. *J Ethnopharmacol.* 2000;72(3):435-41.
76. Dutta H, Paul SK, Kalita D, Mahanta CL. Effect of acid concentration and treatment time on acid-alcohol modified jackfruit seed starch properties. *Food Chem.* 2011;128(2):284-91.
77. Wangchu L, Singh D, Mitra SK. Studies on the diversity and selection of superior types in jackfruit (*Artocarpus heterophyllus* Lam.). *Genet Resour Crop Evol.* 2013;60(5):1749-62.
78. Prakash O, Kumar R, Mishra A, Gupta R. *Artocarpus heterophyllus* (jackfruit): an overview. *Pharmacogn Rev.* 2009;3(6):353-58.
79. Meera M, Ruckmani A, Saravanan R. Anti-inflammatory effect of ethanolic extract of spine, skin and rind of jackfruit peel: a comparative study. *Nat Prod Res.* 2018;32(22):2740-44.
80. Hameed BH. Removal of cationic dye from aqueous solution using jackfruit peel as non-conventional low-cost adsorbent. *J Hazard Mater.* 2009;162(1):344-50.
81. Jadhav VM, Kamble SS, Kadam VJ. Herbal medicine, *Syzygium cumini* review. *Res J Pharm.* 2009;2(8):1212-9.
82. Patel S, Shanmugarajan TS, Somasundaram N, Maity N. Protective effect of *Syzygium cumini* seeds against doxorubicin induced cardiotoxicity in rats. *Int J Pharm Life Sci.* 2010; 1(2):23.
83. Oliver B. Oral hypoglycaemic plants in West Africa. *J Ethnopharmacol.* 1980;2:119-27.
84. Muruganandan S, Srinivasan K, Chandra S. Anti-inflammatory activity of *Syzygium cumini* bark. *Fitoterapia.* 2001;72(4):369-75.
85. Harborne JB. *Phytochemical methods: a guide to modern techniques of plant analysis.* 3rd ed. New York: Chapman and Hall; 1998.
86. Kelen M, Tepe B. Chemical composition, antioxidant and antimicrobial properties of the essential oils of three *Salvia* species from Turkish flora. *Bioresour Technol.* 2008;99:4096-104.
87. Prasad KN, Yang B, Ruenroengklin N. Application of ultrasonication or high-pressure extraction of flavonoids from litchi fruit pericarp. *J Food Process Eng.* 2009;32:828-43.
88. Queiroz ER, Abreu CMP. Constituintes quimicos das fracoes de lichia in natura e submetidas a secagem: potencial nutricional dos subprodutos. *Rev Bras Frutic.* 2012;34(4):1174-9.
89. Bhoopat L. Hepatoprotective effects of lychee (*Litchi chinensis* Sonn.): a combination of antioxidant and anti-apoptotic activities. *J Ethnopharmacol.* 2011;136(1):55-66.
90. Xiang NZ, Kaiwen P, Yan Q, Juxiang W. Effects of the kernel oil from *Litchi chinensis* seed on the level of serum lipids in rats. *Acta Nutr Sin.* 1996;18(2):159-62.
91. Li CG, Zhang SQ, Wang SY. Intervention of litchi nucleus extract fluid in the blood biochemical indexes of model mice with diabetes mellitus induced by alloxan. *J Chin Clin Rehabil.* 2006;10:61-3.
92. Khan MA. Muheet-e-Azam. New Delhi: CCRUM, India Offset Press; 2012. p. 419-20.
93. Nadkarni AK. *Indian materia medica.* Mumbai: Popular Prakashan; 1954. p. 1191-3.
94. Suralkar AA, Rodge KN, Kamble RD, Maske KS. Evaluation of anti-inflammatory and analgesic activities of *Tamarindus indica* seeds. *Int J Pharm Sci Drug Res.* 2012;4(3):213-7.
95. Anonymous. *PDR for herbal medicine.* 2nd ed. Montvale, NJ: Medical Economics Company; 2000. p. 753-4.
96. Anonymous. *The wealth of India: a dictionary of Indian raw materials and industrial products.* Vol. Sp-W. New Delhi: Publications & Information Directorate, CSIR; 1976. p. 114-23.
97. Chatterjee A, Pakrashi SC. *The treatise on Indian medicinal plants.* Vol. 2. New Delhi: Publications & Information Directorate, CSIR; 1992. p. 46-7.
98. Klingel T, Kremer JI, Gottstein VA. A review of coffee by-products including leaf, flower, cherry, husk, silver skin, and spent grounds as novel foods within the European Union. *Foods.* 2020;9:665.
99. Masatte M, Candia A, Ocheng AG. The commercial viability of tamarind (*Tamarindus indica* Linn) fruit based products for improved income among farmers in Northern and Eastern Uganda. *Afr J Food Sci Technol.* 2015;6(6):167-76.
100. Maiti R, Jana D, Das UK, Ghosh D. Antidiabetic effect of aqueous extract of seed of *Tamarindus indica* in streptozotocin-induced diabetic rats. *J Ethnopharmacol.* 2004;92(1):85-91.
101. Djekrif S, Gheribi Z. Application of a statistical design to the optimization of culture medium for alpha-amylase production by *Aspergillus niger* ATCC 16404 grown on orange waste powder. *J Food Eng.* 2006;73:190-7.
102. Kalra P, Sharma S, Suman Kumar S. Antiulcer effect of the methanolic extract of *Tamarindus indica* seeds in different experimental models. *J Pharm Bioallied Sci.* 2011;3(2):236-41.
103. Pimple BP, Kadam PV, Badgujar NS, Bafna AR, Patil MJ. Protective effects of *Tamarindus indica* Linn against paracetamol induced hepatotoxicity in rats. *Indian J Pharm Sci.* 2007;69(6):827-31.
104. Flamini G, Cioni PL, Morelli I. Use of solid-phase micro-extraction as a sampling technique in the determination of volatiles emitted by flowers, isolated flower parts and pollen. *J Chromatogr A.* 2003;998:229-33.
105. Goldhamer DA, Intrigliolo DS. Citrus. In: Steduto P, Hsiao TC, Fereres E, Raes D, eds. *Crop yield response to water.* 1st ed. Rome: Food and Agriculture Organization of the United Nations; 2012. p. 300-15.
106. Chau CF, Huang YL. Comparison of the chemical composition and physicochemical properties of different fibers prepared from the peel of *Citrus sinensis* L. cv. *Liucheng*. *J Agric Food Chem.* 2003;51:2615-8.
107. Haleva-Toledo E, Naim M. Effects of L-cysteine and N-acetyl-L-cysteine on 4-hydroxy-2,5-dimethyl-3(2H)-furanone (furanol), 5-(hydroxymethyl)furfural, and 5-methylfurfural formation and browning in buffer solutions containing either rhamnose or glucose and arginine. *J Agric Food Chem.* 1999;47:4140-5.
108. Castilho LR, Medronho RA. Production and extraction of pectinases obtained by solid state fermentation of agro-industrial residues with *Aspergillus niger*. *Bioresour Technol.* 2000;71:45-50.
109. Kaviya S, Santhanalakshmi J. Biosynthesis of silver nanoparticles using *Citrus sinensis* peel extract and its antibacterial activity. *Spectrochim Acta A Mol Biomol Spectrosc.* 2011;79:594-8.
110. Camacho-Corona MR, Ramirez-Cabrera MA. Activity against drug resistant-tuberculosis strains of plants used in Mexican traditional medicine to treat tuberculosis and other respiratory diseases. *Phytother Res.* 2008;22:82-5.
111. Liu L, Xu. Structure-activity relationship of citrus polymethoxylated flavones and inhibitory effects on *Aspergillus niger*. *J Agric Food Chem.* 2012;60:4336-41.
112. Van-Hung P, Chi PT, Phi NT. Comparison of antifungal activities of Vietnamese citrus essential oils. *Nat Prod Res.* 2013;27:506-8.
113. Xiao H, Yang CS, Li S, Jin H, Ho CT, Patel T. Monodemethylated polymethoxyflavones from sweet orange (*Citrus sinensis*) peel inhibit growth of human lung cancer cells by apoptosis. *Mol Nutr Food Res.* 2009;53:398-406.

114. Sergeev IN, Ho CT, Li S, Colby J, Dushenkov S. Apoptosis-inducing activity of hydroxylated polymethoxyflavones and polymethoxyflavones from orange peel in human lung cancer cells. *Mol Nutr Food Res*. 2007;51:1478-84.
115. Fan K, Kurihara N, Abe S, Ho CT, Ghai G, Yang K. Chemo-preventive effects of orange peel extract (OPE). I: OPE inhibits intestinal tumor growth in ApcMin/+ mice. *J Med Food*. 2007;10:11-17.
116. Kanaze FI, Termentzi A, Gabrieli C. The phytochemical analysis and antioxidant activity assessment of orange peel (*Citrus sinensis*) cultivated in Greece-Crete indicates a commercial source of hesperidin. *Biomed Chromatogr*. 2009; 23:239-49.
117. Kamal G, Anwar F, Hussain A. Yield and chemical composition of Citrus essential oils as affected by drying pretreatment of peels. *Int Food Res J*. 2011; 18(4):1275.
118. Vekari SA, Protopapadakis EE, Papadopoulou P. Composition and seasonal variation of the essential oil from leaves and peel of a Cretan lemon variety. *J Agric Food Chem*. 2002;50(1):147-53.
119. Mohapatra D, Mishra S, Sutar N. Banana and its by-product utilization: an overview. *J Sci Ind Res*. 2010;69:323-9.
120. Dhanavade MJ, Jalkute CB, Ghosh JS. Study antimicrobial activity of lemon (*Citrus lemon L.*) peel extract. *Br J Pharmacol Toxicol*. 2011;2(3):119-22.
121. Mahmoud K. Statistical optimization of cultural conditions of a halophilic alpha-amylase production by halophilic *Streptomyces* sp. grown on orange waste powder. *Biocatal Agric Biotechnol*. 2015;4:685-93.
122. Lohiya NK, Manivannan B. Chloroform extract of Carica papaya seeds induces reversible azoospermia in langur monkey. *Asian J Androl*. 2002;4:17-26.
123. Rumbaoa RGO. Phenolic content and antioxidant capacity of Philippine potato (*Solanum tuberosum*) tubers. *J Food Compos Anal*. 2009;22:546-50.
124. Al-Farsi MA, Lee CY. Optimization of phenolics and dietary fibre extraction from date seeds. *Food Chem*. 2008;108:977-85.
125. Besbes S, Blecker C, Deroanne C, Lognay G, Drira NE, Attia H. Heating effects on some quality characteristics of date seed oil. *Food Chem*. 2005;91:469-76.
126. Said A, Leila A, Kaouther D, Sadia B. Date wastes as substrate for the production of alpha-amylase and invertase. *Iran J Biotechnol*. 2014;12:41-9.
127. Adeosun MA, Oni SO. Phytochemical, minerals and free radical scavenging profiles of Phoenix dactylifera L. seed extract. *J Taibah Univ Med Sci*. 2016;11(1):1-6.
128. Laouini SE, Segni L, Ouahrani MR, Gherraf N, Mokni S. Phytochemical analysis, antioxidant and antimicrobial activities of leaves extract of date palm grown in Algeria. *J Fund Appl Sci*. 2012;4:48-58.
129. Tawfik MS, Saleh EA, Abu-Tarboush HM. Phenolic contents and antioxidant activity of various date palm (*Phoenix dactylifera L.*) fruits from Saudi Arabia. *Food Nutr Sci*. 2011;2:1134-41.
130. Jurenka JS. Therapeutic applications of pomegranate (*Punica granatum L.*): a review. *Altern Med Rev*. 2008;13(2):128-44.
131. Schubert SY, Lansky EP. Antioxidant and eicosanoid enzyme inhibition properties of pomegranate seed oil and fermented juice flavonoids. *J Ethnopharmacol*. 1999;66(1):11-7.
132. Li Y, Guo C. Evaluation of antioxidant properties of pomegranate peel extract in comparison with pomegranate pulp extract. *Food Chem*. 2006;96(2):254-60.
133. Jahfar M, Vijayan K. Studies on a polysaccharide from the fruit rind of *Punica granatum*. *Res J Chem Environ*. 2003; 7:1.
134. Syed DN, Malik A. Photochemopreventive effect of pomegranate fruit extract on UV mediated activation of cellular pathways in normal human epidermal keratinocytes. *Photochem Photobiol*. 2006;82(2):398-405.
135. Aslam MN, Lansky EP. Pomegranate as a cosmeceutical source: pomegranate fractions promote proliferation and procollagen synthesis and inhibit matrix metalloproteinase-1 production in human skin cells. *J Ethnopharmacol*. 2006;103(3):311-8.
136. Olapour S, Mousavi E. Evaluation anti-diarrheal effects of pomegranate peel extract. *J Iran Chem Soc*. 2009;6:115-43.
137. Huang PW, Tsuen HM. Anxiolytic effect of seed of *Ziziphus jujuba* in mouse models of anxiety. *J Ethnopharmacol*. 2000;72(3):435-41.