

Bioprospecting and Therapeutic Applications of *Cocos nucifera* L. Sprouts

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ABSTRACT

Introduction: Cocos nucifera L. is known for all its multiple utilities especially for its nutritional value. The Coconut's basal part of the embryo enlarges to form sprout/haustorium.

Aim: The objective is to compare the phytoconstituents and its bioefficacy of the coconut meat (cellular endosperm) and its sprouts (haustorium) using bioassays leading to novel drug development recommending for the development of nutraceuticals.

Methodology: Studies on toxicity, shelf-life of the dried powdered samples were carried out. Quantification of total lipids, flavonoids, vitamin- C, determination of amylase activity was performed using methanol and aqueous extracts. Bioassays such as antibacterial, *in vitro* antioxidant and anti-inflammatory assays, were conducted. Phytoconstituents were characterized by GC-MS. The cytotoxic effect of the samples was analyzed in human gastric adenocarcinoma epithelial (AGS) cell line infected with gastric ulcer.

Results: Toxicity and shelf-life studies had indicated that the samples were non-toxic with the shelf-life extending up to 180 days. Quantitative analysis showed maximum flavonoids followed by vitamin-C with reduced levels of lipids in the coconut sprouts when compared to coconut meat. Promising amylase activity was revealed by coconut sprouts as these had the potency to decrease the concentration of starch substrate. Prominent zone of inhibitions in the antibacterial assay was recorded in the samples assayed. *In vitro*, antioxidant assays and anti-lipoxygenase activity signified the enriched bioactive potential of the co-conut sprouts. GC-MS analysis had revealed variations in the presence of primary and secondary metabolites. Coconut sprouts extracts were found to be cytotoxic to the infected cells.

Conclusion: Coconut sprouts can be recommended as an economically potent natural product for the commercial production of nutraceuticals in the food industry further to be used as a nutrient supplement in the prevention and management of peptic ulcers with a cost-effective approach.

Key Words: Coconut sprouts, Coconut meat, Phytoconstituents, Phytochemical characterization, Cell line, Nutraceuticals

INTRODUCTION

Coconut (*Cocos nucifera* L.) belonging to Arecaceae is well known for all its utilities. India is the 3rd largest coconut producer where, Tamil Nadu, Kerala, Andhra Pradesh and Karnataka is said to produce 90 per cent of the coconut production. Coconut products are used in daily life by different classes of people. These are some of the prominent sources for the development of medicines and industrial products. Coconut and its products are used as folk medicine and therefore, the coconut palm in Indian classics is known as 'Kalpavriksha'.¹ After the germination of the coconut, the embryo (basal part) usually enlarges and produce sprout/haustorium and its cardioprotective effect was evaluated.² Coconut sprouts wine was produced using *Saccharomyces cerevisiae* which was found to be enriched with antioxidant activity.³Primary and secondary phytoconstituents derived from natural products have gained attention for the prevention and management of several diseases. The presence of squalene, a potent triterpenoid with anti-ulcer activity in the sprouts of *Cocos nucifera* L. was confirmed through *in silico* molecular docking.⁴

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Peptic ulcer occurs in the entire gastrointestinal tract. Gastric ulcers are produced when there is an imbalance between toxic and cytoprotective factors. Toxic endogenous factors include leukotrienes, pepsin, refluxed bile, hydrochloric acid and reactive oxygen species. Toxic exogenous factors include Helicobacter pylori, ulcer-causing gram-negative bacterial strains, steroidal as well as non-steroidal antiinflammatory drugs (NSAIDs), alcohol which initiates the secretion of gastric acid and pepsin.⁵ Cytoprotective factors include prostaglandins, surface-active phospholipids, cell renewal followed by its migration, mucosal blood flow, enzymatic and non-enzymatic antioxidants and mucus-bicarbonate barrier. In recent years, the peptic ulcer has become a major challenge affecting many people. Recent advancements in gastric ulcer therapy aim to suppress the production of gastric acid through the use of antacids, H₂-receptor inhibitors/blockers which include famotidine, ranitidine and several anticholinergics especially telezipine and pirenzepine or sometimes proton-pump inhibitors/blockers such as lansoprazole and omeprazole. However, the major drawback in these advances in modern medicine is the major side effects with limited bioefficacy caused by these drugs.⁶ Thus, in the present scenario, there is a need for prevention and management of peptic ulcers especially gastric ulcers using plant-derived natural products.

Coconut sprouts is a great boon to the food and pharmaceutical industries for the development of nutrient supplements and novel drugs for managing peptic ulcer. Since not much work has been carried out in *Cocos nucifera* L. sprouts, the objective and purpose of the present study are to compare the phytoconstituents and its bioefficacy of the coconut meat and its sprouts using various bioassays leading to novel drug development recommending for the development of nutraceuticals.

MATERIALS AND METHODS

Sample collection

Mature coconut (*Cocos nucifera* L.) and its sprouts (haustorium) were procured from Thirunelveli District, Tamil Nadu, India and were processed by shade dry method.⁷ (Figure 1).

Toxicity analysis

Dried powdered samples were analyzed for the presence of toxic heavy metals such as arsenic, cadmium, cobalt, chromium, mercury, nickel and lead using Inductively Coupled Plasma Optical Emission Spectrometry.⁸

Shelf-life study

Dried powder of the samples collected during the initial period of the study, after 3rd month and 6th month were screened for microbial content using serial dilution technique. Dilutions of

 10^{-6} , 10^{-7} , 10^{-8} and 10^{-9} for total bacterial count and dilutions of 10^{-2} , 10^{-3} , 10^{-4} and 10^{-5} for total fungal count were used to test the microbial quality.⁹

Extraction and Quantificationstudies

Dried powdered samples were extracted with butanol, acetone, chloroform, methanol, aqueous w/v (1:10) using the cold percolation method.⁷Based on qualitative phytochemical studies⁴, methanol and aqueous extracts were taken for the present study. For the documentation of results, sample extracts were abbreviated as, coconut meat extracted with methanol (CCM), aqueous (CCAq); coconut sprouts extracted with methanol (CSM), aqueous (CSAq). Lipids¹⁰, flavonoids¹¹ and vitamin- C¹² were quantified and amylase activity was assayed.¹³

Antibacterial assay

Sample extracts of 20, 40, 60and 80 µg mL⁻¹ were assayed against *Bacillus subtilis, Staphylococcus epidermidis, Escherichia coli* and *Pseudomonas aeruginosa* using agar well diffusion method.⁴

In vitro bioassays

In vitro antioxidant assays [assessment of metal chelating activity,¹⁴ 2,2'-Azino-bis(3-ethylbenzthiazoline-6-sulphonic acid) ABTS⁺ assay]¹⁵ and assessment of anti-lipoxygenase activity¹⁶ (anti-inflammatory assay) was conducted.

Gas Chromatography-Mass Spectrometry (GC-MS) analysis

GC-MS analysis¹⁷ of methanol extracts of the samples were conducted at Sophisticated Analytical Instrument Facility (SAIF), IIT Madras, Chennai, Tamil Nadu, India.

3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay

Human gastric adenocarcinoma (AGS) epithelial cell line with mucus-secreting epithelial cells was obtained from National Centre for Cell Science (NCCS), Pune, India. Indomethacin, an NSAID was used as a gastric ulcer causing agent. Cimetidine, an anti-ulcer drug was used as standard. MTT assay using sample extracts were performed.⁷

Statistical analysis

For each experiment, the data presented are the means of three replicates. Values are expressed as mean \pm SE.

RESULTS

Toxicity analysis

Heavy metals (As, Cd, Co, Cr, Hg, Ni, Pb) were found to be below the detectable limit (BDL) in the samples.

Shelf-life study

The study revealed nil growth of pathogenic bacteria and fungi at different dilutions used thereby indicating that the dried powdered samples (initial period of the study, after 3rd and 6th month) were found to be sterile and free from the microbial load with good shelf-life extending up to 180 days (Figure 2).

Quantificationstudies and determination of amylase activity

Total flavonoid content was reported as Quercetin equivalent, standard curve (0.0097x + 0.1349; $R^2 = 0.9981$). Methanol and aqueous coconut sprouts extracts had maximum flavonoids (83.89 ± 0.07 and 82.37 ± 0.19 mg of QE g⁻¹) whereas coconut meat extracts had 62.25 ± 0.20 and 61.38 ± 0.23 mg of QE g⁻¹ of flavonoids. Vitamin- C was calculated from ascorbic acid standard curve (0.0096x + 0.0427; $R^2 = 0.9986$). Methanol and aqueous coconut sprouts extracts had maximum vitamin- C (0.74 ± 0.02 and 0.73 ± 0.02 mg g⁻¹) whereas, coconut meat extracts had 0.38 ± 0.03 and 0.37 ± 0.02 mg g⁻¹ of vitamin- C. Total lipids were calculated from cholesterol standard curve (0.0098x + 0.041; $R^2 = 0.9966$). Methanol and aqueous coconut sprouts extracts had reduced levels of lipids (0.21 ± 0.02 and 0.25 ± 0.02 mg g⁻¹) whereas, coconut meat extracts had 0.71 ± 0.04 and 0.78 ± 0.03 mg g⁻¹ of total lipids (Figure 3).

Amylase activity monitored over 60 minutes period at 15 minutes interval, indicated that the concentration of substrate decreased with the time reducing from 18.2 ± 0.21 to $2.4 \pm 0.12 \ \mu g \ mL^{-1}$ and 19.1 ± 0.22 to $2.8 \pm 0.11 \ \mu g \ mL^{-1}$ in methanol and aqueous coconut sprout extracts when compared to the coconut meat extracts (20.3 ± 0.22 to $3.1 \pm 0.13 \ \mu g \ mL^{-1}$ and 21.7 ± 0.23 to $4.3 \pm 0.15 \ \mu g \ mL^{-1}$ in 60 minutes). It was observed that there were no marked variations between the extracts whereas remarkable variations were found between the coconut meat and its sprouts indicating that the sprouts are rich in phytoconstituents.

Antibacterial assay

Methanol and aqueous coconut sprouts extracts indicated a maximum zone of inhibitions of 35.66 ± 0.27 mm and 35.37 ± 0.29 mm at 80 µg mL⁻¹ against *Staphylococcus epidermidis* and maximum zone of inhibitions of 34.89 ± 0.27 mm and 34.78 ± 0.25 mm at 80 µg mL⁻¹ against *Pseudomonas aeruginosa*. Coconut meat extracts indicated a zone of inhibitions (28.59 ± 0.28 mm and 27.93 ± 0.23 mm) against *Staphylococcus epidermidis*. Remarkable variations were observed among the sample extracts (Figure 4).

In vitro bioassays

The antioxidant and anti-inflammatory potential of the sample extracts were determined based on the IC_{50} . 100 µg mL⁻¹ showed maximum bioactivities. In metal chelating activity, methanol and aqueous coconut sprouts extracts showed

maximum per cent inhibition (85.22 ± 0.26 and 84.75 ± 0.28) with IC₅₀ values, 33.01 and 35.13 µg mL⁻¹. Methanol and aqueous coconut meat extracts revealed per cent inhibition of 54.57 ± 0.29 and 53.27 ± 0.27 with IC₅₀ values, 87.72 and 90.35 µg mL⁻¹. Na₂EDTAindicated 86.24 ± 0.26 per cent inhibition with IC₅₀ value, 31.0 µg mL⁻¹. ABTS⁺ assay revealed that methanol and aqueous coconut sprouts extracts showed maximum per cent inhibition (88.39 ± 0.27 and 87.32 ± 0.27) with IC₅₀ values, 31.29 and 32.96 µg mL⁻¹. Methanol and aqueous coconut meat extracts revealed per cent inhibition of 52.66 ± 0.27 and 51.54 ± 0.29 with IC₅₀ values, 96.67 and 99.88 µg mL⁻¹. Ascorbic acid indicated 90.34 ± 0.26 per cent inhibition with IC₅₀ value, 28.68 µg mL⁻¹.

Concerning anti-inflammatory assay, methanol and aqueous coconut sprouts extracts showed maximum per cent inhibition (92.27 \pm 0.25 and 91.78 \pm 0.24) with IC₅₀ values, 26.66 and 27.90 µg mL⁻¹. Methanol and aqueous coconut meat extracts revealed per cent inhibition of 62.97 \pm 0.23 and 61.88 \pm 0.21 with IC₅₀ values, 68.69 and 71.07 µg mL⁻¹. Indomethacin indicated 94.46 \pm 0.27 per cent inhibition with IC₅₀ value, 21.85 µg mL⁻¹. There were marked variations between the coconut meat and its sprouts, in which the sprout extracts had maximum antioxidant and anti-inflammatory activity on par with standard (Figure 5).

Gas Chromatography-Mass Spectrometry (GC-MS) analysis

Various peaks were observed in coconut sprouts extract representing the volatile compounds of the phytoconstituents such as flavonoids, terpenoids, phenols, proteins, amino acids, carbohydrates, vitamins, essential fatty acid esters when compared to the coconut meat which had phenols, alkaloids, fatty acid esters (Figure 6). 3',4',5,7-Tetrahydroxyflavone was found to be abundantly present in coconut sprouts with a peak area per cent of 88.2 (Table 1 & Table 2).

MTT assay

Methanol and aqueous coconut sprouts extracts (100 µg mL⁻ ¹) showed maximum per cent inhibition of infected AGS cells (93.92 \pm 0.23 and 92.12 \pm 0.27) with IC $_{50}$ values, 20.23 and 22.11 µg mL⁻¹. Methanol and aqueous coconut meat extracts revealed a per cent inhibition of 67.11 ± 0.25 and 66.12 \pm 0.26 with IC₅₀ values, 67.66 and 68.98 µg mL⁻¹. Cimetidine $(100 \ \mu g \ mL^{-1})$ indicated 94.85 ± 0.29 per cent inhibition with IC₅₀ value, 18.83 µg mL⁻¹. Methanol and aqueous extracts of the coconut sprouts were found to be cytotoxic to the infected AGS cells with gastric ulcers in a dose-dependent manner when compared to the coconut meat extracts (Figure 7). An increase in the concentration of sample extracts has increased the cytotoxic activity of the samples. There was a remarkable variation between the samples in which coconut sprouts extracts had maximum anti-ulcer activity on par with the standard.

DISCUSSION

Bioprospecting of natural plant-derived food products leads to the study of phytoconstituents and the identification of novel compounds with pharmaceutical properties.

Plant and plant-derived food products may be contaminated with toxic heavy metals or other residues resulting in health issues. Lead, cadmium, mercury, chromium in the food products cause gastric ulcer/cancer through the disruption of the gastric mucosa. These heavy metals along with arsenic were reported to be exogenous sources of ROS resulting in gastric mucosal tissue damage, epithelial, endothelial inflammation, where, cadmium affect E-cadherin function. Cobalt induces DNA damage, nickel modify the activity of catalase through ROS generation causing gastric ulcer/cancer.¹⁸ Hence, it is essential to analyze the toxicity of the samples. ICP-OES analysis was applied to metal concentrations in the samples. Present findings are clear evidence that the samples were found to be non-toxic and can be recommended for consumption as a natural edible product for the prevention and management of several diseases.

During germination, moisture content, nutrients released and the warm appropriate temperature are favourable for the growth of pathogens. Analyzing the microbiological quality of the samples signifies the importance of the study. The present study proves that the dried powdered samples were found to be pure and free from the microbial load with shelflife extending up to 180 days. This reveals that the samples were found to be from a clean environment which can be best used for the preparation of nutraceuticals.

Flavonoids are potent bioactive compounds with a polyphenolic structure occurring in various natural edible plant-derived food products with health-promoting benefits having nutraceutical and pharmaceutical applications. These tend to possess biochemical, antioxidant, anti-inflammatory, anticancer activities due to which the use of flavonoids in the commercial production of nutraceutical and pharmaceutical drugs are in increasing demand. Several reports have indicated that flavonoids possess gastroprotective, cytoprotective and anti-secretory activities.¹⁹ Ascorbic acid or vitamin-C, a water-soluble vitamin is responsible for reducing the bleeding in gastric ulcers, in addition to the reduction of the NSAIDs related gastric mucosal destructions or damage and Helicobacter pylori suppression. Lipids include fats, phospholipids, sterols and glycolipids which are almost found in all living cells. Glycolipids are found in edible parts of the plant. Increased lipid content may lead to gastric ulcers and carcinomas. The present finding is scientifically validated that the coconut sprouts are enriched bioactive compounds and reduced level of lipids than the coconut meat. Thus, coconut sprouts are an alternative source of coconut meat with high cholesterol. It has solved the issue as a natural edible product found to be enriched with potent phytoconstituents used in the prevention and management of ulcers and other diseases with no side effects. Elevated levels of amylase can cause acute gastritis. Amylase activity in the coconut sprouts was found to be at a permissible level and it helps in the breakdown of complex substances. Hence, it is considered safe for different age groups of people to consume coconut sprouts.

Developing resistance breaking natural antimicrobial agents are the need of the hour in the present scenario. Current findings prove that when compared to the coconut meat, the coconut sprouts with the prominent shelf-life period, enriched flavonoids, ascorbic acid act as a natural antibacterial agent. Bioactive flavonoids act against the pathogenic bacteria by inhibiting the synthesis of nucleic acids, functions of the cytoplasmic membrane, energy metabolism resulting in potent antibacterial activity.²⁰

In the metal chelating assay, ferrozine chelate with Fe₂⁺ and form a complex. ROS induce gastric mucosal damage.²¹ Antioxidants in the sample extracts form a coordinate complex with metal ions, inhibit electron transfer and no free radicals are produced. Suppression of ABTS⁺ ions is essential to reduce the release of free radicals resulting in healthy tissue damage. Flavonoids and essential phytoconstituents in the coconut sprouts might be responsible for this antioxidant activity. The present study has proved that when compared to coconut meat extracts, the coconut sprouts had a maximum antioxidant activity with minimal IC_{so} value.

The inflammatory process involves the activity of inflammatory mediators (ROS, nitric oxide, neutrophil-derived free radicals, cytokines and prostaglandins). Increased levels of these mediators lead to injury of the tissues by damaging the macromolecules, lipid peroxidation of the membrane. Lipoxygenases contain non-heme iron dioxygenases mostly involved in leukotrienes synthesis which are essential for inflammation and hence these are the most promising inflammatory target.²² Present findings have proved that the antilipoxygenase activity was found to be in maximum in the coconut sprouts than the coconut meat which indicated that the sprouts are natural bioactive anti-inflammatory agents enriched with essential flavonoids.

In GC-MS analysis, coconut sprouts have maximum flavonoids constituting flavanones, flavonols, flavones, anthocyanidins and isoflavones indicating a promising role in preventing/healing peptic ulcers leading to a cytoprotective shield in the gastrointestinal region. It can be an alternative for the reduction of peptic ulcers associated with *Helicobacter pylori* infection/NSAIDs⁷ thereby contributing therapeutic biological effects.

Human gastric adenocarcinoma (AGS) cell line with mucussecreting epithelial cells used as an *in vitro* model is an appropriate cell line related to peptic ulcers/gastric cancers. Indomethacin (NSAID) is considered an instant gastric ulcer-causing agent which inhibits the synthesis of prostaglandins and thereby causes gastric ulcers. Prostaglandins are protective factors that prevent mucosal damage. Cimetidine, a modern commercial anti-ulcer drug heals gastric ulcers. Decreasing absorbance in the cells treated with the methanol and aqueous extracts of the coconut sprouts suggests cytotoxicity to AGS infected cells with gastric ulcers. MTT assay proved that the coconut sprouts were found to be cytotoxic to the infected AGS cell line with a gastric ulcer in a dosedependent manner. The minimum IC_{50} value of the coconut sprouts extracts has indicated that the sprouts are possessing promising bioactive potential due to enriched phytoconstituents which are involved in various metabolic pathways leading to the inhibition of ulcers.

Significant findings and novelty of the present study include (i) First report of bioprospecting study of sprouts of *Cocos nucifera* L. (haustorium) revealing potent anti-ulcer activity through *in vitro* analysis using AGS cell line; (ii) Presence of flavonoids proves that coconut sprouts can be recommended as a natural edible product; (iii) Commercial production of nutraceuticals for the prevention and management of peptic ulcers; (iv) Presence of specific secondary metabolites in coconut sprouts are found to be advantageous over coconut meat; (v) Hesitation in consumption of coconut meat due to its rich cholesterol and lipid content finds the consumption of coconut sprouts with less cholesterol as an alternative source.

CONCLUSION

A bioprospecting study revealed that the coconut sprouts are enriched with maximum flavonoids, vitamin- C and permissible amylase activity with minimum lipids when compared to the coconut meat. *Cocos nucifera* L. sprouts possess promising antibacterial, antioxidant and anti-inflammatory activities. GC-MS and MTT assay indicated the presence of essential bioactive compounds responsible for the cytotoxic activity of the sprouts against the AGS cell line with gastric ulcer. Thus, coconut sprouts can be recommended as an economically potent natural product for the commercial production of nutraceuticals in the food industry. Further, it can be used as a nutrient supplement in the prevention and management of gastric ulcers with a cost-effective approach as well can be recommended to the pharmaceutical industries for novel drug development and drug discovery.

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Author's Contribution

S. Abiraami Valli performed the experiments, drafted the manuscript, analysed and interpreted the data.

Dr. S. Uma Gowrie, proposed the concept, designed the experiments, supervised, analysed, interpreted the data, technically supported, critically revised and finally approved the manuscript.

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Compound Name Retention Molecular Molecular Peak Compound **Biological activities** time (mins) formula weight (g area % nature (PubChem, NIST & **mol**⁻¹) ChEBI) Quinoline C H N 129.16 66.34 Alkaloid Anti-malarial agent. 10.70 Methyl beta-D-galactopyr-C₇H₁₄O₆ Monosaccharide Anti-tumor agent. 11.63 194.18 54.33 ranoside Dodecanoic acid. C, H, O 456.7 Glyceryl diester Anti-ageing, anticancer 24.727 3.39 1-(Hydroxymethyl)-1,2agent, lubricant. Ethanediyl ester Dodecanoic acid, 2-hy-C15H30O4 Dodecanoate ester Antiviral, antimicrobial, 27.003 65.0 274.4 droxy-1-(hydroxymethyl) emulsifying agent, emolethyl ester lient. Glycerol Tricaprylate 27.878 $C_{77}H_{50}O_{6}$ Triglyceride Anticonvulsant. 470.7 5.59 Nonane, 1-Iodo-C_HI Essential oil Flavouring agent. 29.429 254.15 10.55 Dodecanoic acid, 1,2,3-Pro-C_10H_106 8.80 Glycerol derivative Thickening agent. 639.0 29.534 panetriyl ester Tocopherols C_0H_0O_ Vitamin E Antioxidant. 30.88 416.7 64.72 Lauric anhydride $C_{24}H_{46}O_{3}$ 382.6 68.33 Dodecanoic acid Antimicrobial, anti-ageing 31.705 derivative agent.

Table 1: GC-MS analysis of methanol extract of Cocos nucifera L. meat

Table 2: GC-MS analysis of methanol extract of Cocos nucifera L. sprouts

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Compound Name	Reten- tion time (mins)	Molecular formula	Molecular weight (g mol ⁻¹)	Peak area %	Compound nature	Biological activities (PubChem & NIST)
1,2,4-Triazine-5-thiol, 3-amino-6-methyl-	11.6	$C_4H_6N_4S$	142.19	61.9	Triazole	Anticancer, anticonvulsant, anti- inflammatory, anti-HIV, antituber- cular agent.
Pent-1-yn-3-ene, 4-methyl- 3-phenyl-	14.08	$C_{12}H_{12}$	156.22	8.1	Enyne deriva- tive	Anti-inflammatory, anticancer agent.
Flavone	14.97	$C_{15}H_{10}O_{2}$	222.24	72.3	Flavonoid	Antimicrobial, antiviral, anti-in- flammatory, antioxidant, antidiar- rheal, anti-ulcer agent.
Glucose	15.82	$C_{6}H_{12}O_{6}$	180.16	3.4	Monosaccha- ride	Energy source.

Table 1: (Continued)

Compound Name	Reten- tion time (mins)	Molecular formula	Molecular weight (g mol ⁻¹)	Peak area %	Compound nature	Biological activities (PubChem & NIST)
3',4',5,7-Tetrahydroxyflavone	16.87	$C_{_{15}}H_{_{10}}O_6$	286.24	88.2	Flavonoid	Exhibit antioxidant, anti-diabetic, anti-inflammatory, antiviral, anti- microbial, anticancer, anti-ulcer, anti-allergic activities.
Carbamic acid, phenyl-, phenylmethyl ester	17.47	C ₁₄ H ₁₃ NO ₂	227.26	44.8	Phenol	Anti-tubercular agent.
Quinoxaline, 2-isopropyl- 3-phenyl-, 4-oxide	18.45	$C_{_{17}}H_{_{16}}N_{_2}o$	264.32	54.2	Quinoxaline	Possess anti-tubercular, antiviral, anti-protozoan, anti-parasitic, anti-inflammatory, anti-diabetic activities.
Ascorbic acid	18.45	$C_6H_8o_6$	176.12	69.3	Vitamin-C	Antioxidant, detoxicant.
16-Octadecenoic acid, me- thyl ester	19.2	$C_{19}H_{36}O_{2}$	296.49	47.4	Methyl ester	Exhibit cancer preventive, anti- inflammatory, anti-androgenic, dermatitigenic, properties.
Pyrimidine, 5-ethyl-2-[4-(4- ethylcyclohexyl)-phenyl]-	19.88	$C_{20}H_{26}N_{2}$	294.4	14.0	Pyrimidine group	Anti-inflammatory, antiviral, an- tipyretic, antioxidant, anticonvul- sant, analgesic, antihypertensive, anti-diabetic, anticancer agent.
Squalene	21.08	$C_{30}H_{50}$	410.7	68.5	Triterpenoid	Anticancer, anti-inflammatory agent.
Phenol, 2,6-bis(1,1- dimethylethyl)-4-[(4-hy- droxy-3,5-dimethylphenyl) methyl]-	22.45	$C_{23}H_{32}O_{2}$	340.5	51.4	Polyphenol	Antioxidant.
3,4-Dihydroxy-1,6-bis-(3- methoxy-phenyl)-hexa-2,4- diene-1,6-dione	24.23	$C_{20}H_{18}O_{6}$	354.4	64.7	Diarylhepta- noid	Hepato-protective, anti-cathartic, anti-ulcer, anti-diabetic, diuretic, anti-neoplastic agent and lowers cholesterol.



Figure 1: (a) & (b) Coconut meat and sprout (c) & (d) Dried powder of coconut meat and its sprouts.



Figure 2: Total Bacterial & Total Fungal Count (after 6th month) (a) & (b) Coconut meat (c) & (d) Coconut sprouts.



Figure 3: (a) & (b) Total Flavonoid, vitamin- C, lipid content in the sample extracts.



Figure 4: (a) & (b) Methanol and aqueous sprout extracts against *Staphylococcus epidermidis* (c) & (d) Methanol and aqueous sprout extracts against *Pseudomonas aeruginosa* (e) Anti-bacterial activity of the sample extracts.



Figure 5: *In vitro* antioxidant and anti-inflammatory activity of the sample extracts.



Figure 6: (a) & (b) GC-MS spectrum of methanol extract of the samples.



Figure 7: Illustration of cytotoxic activity (a) Control (b) Standard (c) & (d) Methanol and aqueous coconut meat extract (e) & (f) Methanol and aqueous coconut sprouts extract (g) Cytotoxic MTT assay of the sample extracts.