Ethidium Bromide-Agar Cartwheel Method in the Detection of Efflux Pump Mediated Multi-Drug Resistance in Enterobacteriaceae

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ABSTRACT

Introduction: Multidrug-resistant (MDR) strains of bacteria pose a major threat in clinical settings. Multidrug resistance can be due to various mechanisms but is primarily the result of over-expressed efflux pumps that extrude unrelated antibiotics before they reach the intended targets. The emergence of MDR due to efflux pumps has led to a diagnostic and therapeutic dilemma. Ethidium Bromide (EtBr)-agar cartwheel assay is a newly discovered simple, safe and cost-effective method to determine efflux pump activity.

Objectives: The study aimed at the detection of efflux pump activity in MDR strains belonging to Enterobacteriaceae family using the EtBr cartwheel method as well as determining the antimicrobial susceptibility pattern of MDR Enterobacteriaceae.

Materials and methods: A total of 95 MDR Enterobacteriaceae isolates from various clinical samples were included in the study. Identification and antimicrobial susceptibility were done following Clinical Laboratory Standards Institute (CLSI) 2019 guidelines. They were evaluated for efflux activity using the EtBr agar cartwheel method. Data analysis was performed using SPSS version 20.

Results: All the 95 isolates were tested for efflux pump using the Ethidium Bromide agar cartwheel technique. The results showed that 47 (49.47%) of the total isolates gave positive results. Among the 47 efflux pump, positive organisms majority were Klebsiellapneumoniae, followed by Escherichia coli and Proteus mirabilis.

Conclusion: Laboratory detection of efflux pumps in bacteria can be effectively done using EtBr agar cartwheel assay. We conclude that over-expression of efflux pumps has led to an alarming rise in drug resistance and necessary steps should be taken to control this problem.

Key Words: Ethidium Bromide, Cartwheel, Efflux Pump, Multidrug resistance, Enterobacteriaceae, Klebsiellapneumoniae

INTRODUCTION

Bacterial infections have again become a threat due to the rapid emergence of resistant bacteria caused by the overuse and misuse of antibiotics, thus endangering their efficacy. Bacteria resist the action of antibiotics through several mechanisms.¹ Out of these, bacterial efflux pumps are becoming a major concern because they provide bacteria with the ability to drive away many structurally unrelated antibiotics, even before their effect begins to onset.² These pumps are classified into five: ATP binding cassette superfamily (ABC), major facilitator superfamily (MFS), resistance nodulation cell division superfamily (RND), small multidrug resistance family (SMR), multi-antimicrobial extrusion protein family (MATE).³⁴⁵

The intestinal tracts of humans and animals form the natural habitat for Enterobacteriaceae which are a heterogeneous group of Gram-negative rods. Enterobacteriaceae cause a variety of human infections that can be broadly classified as either enteric diseases or extra-intestinal infections such as urinary tract infections, bacteraemia, and meningitis. Enterobacteriaceae acquired in the hospital environment are often resistant to many antimicrobial agents.⁶

The evaluation of efflux systems by conventional methods such as the retention of the fluorescent dye ethidium bromide (EtBr) or radio-labelled antibiotics requires specialised instrumentation not usually available in a clinical laboratory. Hence there is a need to develop a fast and cost-effective...
method for detecting efflux pumps in efflux mediated multi-drug-resistant (MDR) bacteria. The EtBr agar cartwheel assay is a newly discovered simple, instrument-free, safe and cost-effective method utilised for the demonstration of efflux pump activity in bacteria. It employs EtBr as the pump substrate that allows the verification of the existence of an over-expressed efflux system. Thus, there is a maximum concentration of EtBr which is effectively extruded by the cells and higher EtBr concentrations will be retained; hence when the bacterial mass is exposed to ultraviolet (UV) light, fluorescence will be detected. The concentration of EtBr that is required to produce fluorescence in bacterial strains over-expressing efflux systems is considerably higher than that which produces fluorescence of the reference strain.\(^5\)\(^7\) The present study aimed to detect efflux pumps in MDR bacteria belonging to the Enterobacteriaceae family with the help of the EtBr agar Cartwheel assay in a tertiary care centre.

**MATERIALS AND METHODS**

This cross-sectional observational study was carried out in the Department of Microbiology, Bangalore Medical College & Research Institute (BMCRI), Bengaluru from samples sourced from the attached hospitals. The sample size was taken as 95 as calculated with \(d(\text{absolute precision})=8\) and \(p(\text{prevalence})=83\%\), using the formula: \(N=4pq/d^2\).\(^5\)\(^7\) MDR Enterobacteriaceae isolates from clinical samples sent for culture and antimicrobial susceptibility testing were included in the study. Susceptible Enterobacteriaceae, Gram-negative organisms other than Enterobacteriaceae and Gram-positive cocci were excluded from the study. Institutional ethical clearance was obtained. Demographic details like name, age, gender, place and other relevant medical details were obtained from request forms sent to the microbiology laboratory and from the medical records department.

Identification and anti-microbial susceptibility were done using VITEK 2 system as per Clinical Laboratory Standards Institute (CLSI) 2019 guidelines.\(^5\)\(^7\) MDR isolates were further tested for efflux activity using the Ethidium Bromide Agar Cartwheel method. Escherichia coli (E.coli) ATCC25922 was used as control.

**EtBr-agar cartwheel method:**

Bacterial strains were grown in 5 mL of appropriate liquid broth until they reached an optical density (OD) 0.6 at 600 nm. The OD of the cultures was adjusted with PBS to 0.5 McFarland standard. Tryptic soy agar plates containing EtBr concentrations ranging from 0 to 2.5 mg/L were prepared on the same day of the experiment and protected from light. The plates were then divided into as many as 12 sectors by radial lines (cartwheel pattern) as exemplified in Flowchart 1. OD adjusted cultures were inoculated on EtBr-agar plates starting from the centre of the plate and spreading towards the edges, as indicated by the arrowheads shown in Flowchart 1. Each plate included at least one reference strain that served as a comparative control. The number of reference strains to be included may be increased to two or more, depending on a given experiment. The swabbed EtBr-agar plates were then incubated at 37°C for 16 hours and examined under a suitable source of UV light, such as a hand-held UV lamp or a UV transilluminator. The minimum concentration of EtBr (MCEtBr) that produced fluorescence of the bacterial mass was recorded(Fig 1).\(^5\)\(^7\) The absence of fluorescence determines the presence of active efflux pumps in MDR strains.

**RESULTS**

A total of 95 Enterobacteriaceae organisms were isolated from various specimens of patients, of which 54(56.8%) were Klebsiella pneumoniae (K.pneumoniae), 27(28.4%) were E.coli, 8(8.4%) were Proteus mirabilis (P.mirabilis), 3(3.2%) were Enterobacter cloacae, 1(1.1%) was Enterobacter aerogenes, 1(1.1%) was Providencia stuartii and 1(1.1%) was Providencia rettgeri. (Table 1)

83 isolates were recovered from inpatients and 12 isolates were from the outpatient’s department. The distribution of the patients included in the study among various departments was as follows: 26(27.37%) in Surgical units, 15(15.79%) in Burns ward, 9(9.47%) in Neonatal Intensive Care Unit (ICU), 7(7.37%) in Paediatric units, 6(6.32%) in Obstetrics & Gynaecology (OBG) units, 5(5.26%) in Medicine units, 3(3.16%) in Dermatology units, 2(2.1%) each in Medical ICU, Burns ICU, Pulmonary Medicine units and Paediatric ICU, 1(1.05%) each in Psychiatry, Otorhinolaryngology, OBG ICU, Urology and Orthopaedic units. 10isolates (10.57%) were not traceable.

Specimens included in the study were: pus 48 (50.5%), urine 18 (18.9%), endotracheal aspirate 13(13.7%), Blood 5(5.3%), Sputum 5(5.3%), CSF 2(2.1%), High vaginal/cervical swab 2(2.1%) and fluid aspirates2 (2.1%).

**Antimicrobial susceptibility pattern**

Antibiotic susceptibility to various antimicrobials was done by automated identification/ Antimicrobial susceptibility (ID/AST) method using VITEK-2 (Biomerieux). All 95 (100%) isolates were resistant to cefuroxime, cefuroxime
axetil and ceftriaxone. 90 isolates (94.7%) were resistant to ciprofloxacin, followed by resistance to piperacillin/tazobactam (93.68%), amoxicillin/clavulanic acid (91.58%) and cefoperazone/sulbactam (85.26%). Least resistance was noted among the isolates to nitrofurantoin (6.3%) and colistin (8.4%). (Fig 2)

**The mean age of patients was 33.21 ± 24.216, M: F ratio was 12:7.**

*E. coli* (27) isolates were most resistant to 2nd and 3rd generation cephalosporins (100%) followed by 4th generation cephalosporins and fluoroquinolones (96.23%) followed by ampicillin (92.56%) with the least resistance to tigecycline and nitrofurantoin (0%).

*K. pneumoniae* (54) isolates were also most resistant to 2nd and 3rd generation cephalosporins and amoxicillin/clavulanic acid (100%) followed by piperacillin/tazobactam (98.15%) and cefoperazone/sulbactam (94.44%). Colistin resistance was lower among these isolates (3.7%).

*P. mirabilis* (8) isolates were most resistant to 2nd and 3rd generation cephalosporins, amoxicillin/clavulanic acid, imipenem, amikacin, gentamicin and ciprofloxacin (100%) followed by ampicillin (87.5%) and trimethoprim/sulphamethoxazole (75%). They were least resistant to nitrofurantoin (0%) and tigecycline (12.5%).

The organisms were also flagged for various mechanisms of resistance by VITEK-2. 75 (78.94%) isolates were flagged for carbapenemase. 63 (66.31%) showed impermeability to carbapenems. 19 (20%) were ESBL producers, 4 (4.21%) showed impermeability to cephems and 1 (1.05%) isolate showed acquired cephalosporin activity. (Table 2)

**Efflux pump activity:**

All the 95 isolates were tested for efflux pump using the Ethidium Bromide-agar cartwheel technique. The results showed that 47 (49.47%) isolates gave positive results out of which 6 did not show fluorescence even at 2.5mg/L (max concentration of EtBr used), 1 isolate did not show fluorescence at 2mg/L but fluoresced at 2.5mg/L and 40 did not fluorescent at 1mg/L only but fluoresced at 2 and 2.5mg/L. (Table 3)

The mean age of the patients who had infections due to efflux pump positive organisms was 34.02± 25.010 and the male: female (M: F) ratio was 30:17.

Among the 47 efflux pump positive organisms 26 (55.32%) were *K. pneumoniae*, 13 (27.66%) were *E. coli*, 5 (10.67%) were *P. mirabilis*, 2 (4.25%) were *Enterobacter cloacae* and 1 (2.13%) were *Enterobacter aerogenes* (Table 1). The efflux pump positive isolates were mostly from pus (56.45%), followed by urine (19.15%).

13 (27.7%) of the efflux pump positive isolates were from surgical cases and 7 (14.9%) were from the burns ward.

38 (80.85%) isolates were recovered from inpatients and 9 (19.15%) isolates were from the outpatient department. 9 (19.15%) isolates were from ICU.

Among the efflux pump positive organisms, some of the organisms were flagged for other mechanisms of resistance like carbapenemase, impermeability to carbapenems, ESBL and impermeability to cephalams, by Vitek-2. (Table 2)

The most commonly used antimicrobials among the patients whose specimens yielded efflux pump positive isolates were ceftriaxone, piperacillin/tazobactam and amikacin.

**DISCUSSION**

Management of infections is very challenging due to the emergence of multidrug resistance. There are very few available options among anti-microbial agents against MDR Gram-negative bacteria, thus posing a major public health threat. Since MDR is primarily the result of over-expressed efflux pumps that extrude unrelated antibiotics before they reach the intended targets, clinical laboratories should develop and implement new and improved methods for the timely identification of efflux mediated MDR phenotypes.

95 organisms were tested for efflux pump activity, which is very high as compared to studies conducted by Rana T et al. and Al Fayyadh Z et al. and Martins M et al.⁵⁻⁷⁻¹⁰

**A. Efflux Pump Activity:-**

In the present study, EtBr fluorescence was not observed in 47/95 (49.47%) MDR isolates, which suggested that these isolates contain efflux pump which effluxed out EtBr from the bacterial cell. 6 isolates showed over-expressed efflux systems by not showing fluorescence even at 2.5mg/L (highest concentration of EtBr used), 1 isolate showed intermediate efflux activity by showing fluorescence at 2.5mg/L but not at 2mg/L and 1mg/L while 40 showed mild efflux pump activity by showing fluorescence at 2 and 2.5mg/L.

In a study conducted by Martins M et al., 42 clinical isolates with a confirmed MDR phenotype were evaluated for efflux pump activity by the EtBr cartwheel method. The study included 10 *Escherichia coli*, 18 *Enterobacter aerogenes*, 10 *Staphylococcus aureus*, and 4 *Enterococcus faecalis* strains. The study findings revealed a presence of efflux activity in 36% of the isolates, which is comparable to the results of our study. Among the efflux pump positive organisms, maximum efflux activity was shown by *E. coli* (16.6%) species, whereas in our study *K. pneumoniae* exhibited maximum efflux activity (27.66%).⁷

In a study conducted by Al Fayyadh Z et al.¹⁰ which included 165 specimens from different sources, 93 isolates were identified as *E. coli*. About 40 *E. coli* isolates were tested for the presence of efflux pump using the cartwheel method.
31 isolates (77.5%) in this study revealed positive results. In our study, the efflux pump activity was relatively lower (49.47%), the reason for which could be the presence of other mechanisms of multidrug resistance.

All 47 efflux pump isolates in the present study were noted to possess one or more of the other mechanisms of resistance: carbapenemase, ESBL, impermeability to cephamycins and impermeability to carbapenem, as revealed by Vitek-2. The other 48 isolates which did not show efflux pump activity also flagged for various other mechanisms of resistance mentioned above along with acquired cephalosporins.

A study by Suwantarat N et al. shows that one of the major contributors to anti-microbial resistant bacteria in southeast Asia is MDR Gram negative bacteria. Overuse of carbapenem therapy to treat these infections has led to the high prevalence of ESBLs in this region. To control the spread of MDRGNs in this region, it is pertinent to improve the infection control practices, have better laboratory detection facilities and advocate judicious use of anti-microbial agents.

B. Anti-microbial susceptibility pattern among Enterobacteriaceae:

In our report, K. pneumoniae (28.4%) is the most common etiological agent of MDR infections followed by E. coli (8.4%). These organisms were most commonly isolated from pus (50.5%) and urine (18.9%) samples. The majority of them were obtained from Surgery (27.37%) and Burns department (15.79%).

But in a study conducted by Beyene D et al. in which 94.5% of the isolates were MDR, E. coli was the most common etiological agent followed by K. pneumoniae. The majority of the isolates were from urine (62.5%) and by blood (28.4%) and 73% were from ICU.

The mean age of patients in our study was 33.21 ± 24.216 years and the male: female ratio was 12:7. But in a study conducted by M.A. Rajiet al. the mean age was 42.4 years with an insignificant difference between the isolates collected from males and females.

These differences could be due to variations in geographic areas, periods of study, target population and sample size.

The isolates encountered in the present study were most resistant to second and third-generation cephalosporins, fluoroquinolones, piperacillin/tazobactam and least resistant to colistin, tigecycline and amikacin (Fig 2). These results are consistent with the results of a study conducted by Charan et al. where the organisms were most resistant to amoxicillin/clavulanic acid, 2nd and 3rd cephalosporins and carbapenems.

The antimicrobial susceptibility findings related to E. coli and K. pneumoniae are consistent with a study conducted by Lai CC et al.

CONCLUSIONS

Laboratory detection of efflux pumps in bacteria can be effectively done using EtBr cartwheel assay. This is a simple and instrument-free technique that can be performed in most laboratories. Based on the present study, it can be concluded that overexpression of efflux pumps has led to an alarming rise in multi-drug resistance and necessary steps should be taken to control this problem. This is expected to aid in controlling hospital-acquired infections and advocating rational use of antimicrobials.

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Conflict of Interest: Nil

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Figure 1: Ethidium bromide agar- cartwheel method depicting efflux pump activity.
Note: Isolate 13 shows no fluorescence at concentration of 1mg/L and 2mg/L and minimal fluorescence at 2.5mg/L as compared to other isolates. This indicates that isolate 13 contains efflux pumps.

| Table 1: Distribution of Organisms Among the Isolates |
|---|---|---|
| Organism | Efflux Pump negative | Efflux Pump positive |
| Klebsiella | 28 (58.33%) | 26 (55.32%) |
| E.coli | 14 (29.17%) | 13 (27.66%) |
| Proteus mirabilis | 3 (6.25%) | 5 (10.64%) |
| Enterobacter cloaceae | 1 (2.08%) | 2 (4.25%) |
| Enterobacter aerogenes | 0 | 1 (2.13%) |
| Providencia stuartii | 1 (2.08%) | 0 |
| Providencia retgeri | 1 (2.08%) | 0 |
| Total | 48 (100%) | 47 (100%) |

Table 2: Other Mechanisms Of Resistance

<table>
<thead>
<tr>
<th>Other mechanisms of MDR</th>
<th>Number of Efflux pump positive organisms (n=47)</th>
<th>Number of Efflux pump negative organisms (n=48)</th>
</tr>
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<tbody>
<tr>
<td>Aquiredcephalosporinase</td>
<td>0/47 (0%)</td>
<td>1/48 (2.08%)</td>
</tr>
<tr>
<td>Carbapenamase</td>
<td>38/47 (80.85%)</td>
<td>37/48 (77.08%)</td>
</tr>
<tr>
<td>ESBL</td>
<td>10/47 (21.28%)</td>
<td>9/48 (18.75%)</td>
</tr>
<tr>
<td>Impermeability to cephemycins</td>
<td>3/47 (6.38%)</td>
<td>1/48 (2.08%)</td>
</tr>
<tr>
<td>Impermeability to carbapenem</td>
<td>30/47 (63.83%)</td>
<td>33/48 (68.75%)</td>
</tr>
</tbody>
</table>

Table 3: Efflux Pump Activity among the Isolates

<table>
<thead>
<tr>
<th>Number of isolates</th>
<th>Concentration of Et-Br at which the bacteria did not fluoresce(mg/L)</th>
<th>Efflux activity</th>
<th>MC Et-Br (MDR)</th>
</tr>
</thead>
<tbody>
<tr>
<td>48</td>
<td>&lt;1</td>
<td>0</td>
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</tr>
<tr>
<td>40</td>
<td>1</td>
<td>+</td>
<td>2</td>
</tr>
<tr>
<td>1</td>
<td>2</td>
<td>++</td>
<td>2.5</td>
</tr>
<tr>
<td>6</td>
<td>2.5</td>
<td>+++</td>
<td>&gt;2.5</td>
</tr>
</tbody>
</table>

Figure 2: Antimicrobial Susceptibility Pattern Among All Isolates.