INTRODUCTION

It is known diabetes can lead to potential complications such as encephalopathy, heart disease, hypertension, tachycardia, kidney damage, stroke, nerve damage, visual impairment, seizures, and other pathologies.1,2 So, there is a growing need for multifunctional drugs to treat diabetes and improve its adverse metabolic effects. In recent years, special attention has been given to the treatment with phytoecdysteroids, as these substances have shown to have a wide range of positive effects on different metabolic pathways. Phytoecdysteroids are involved in the regulation of carbohydrates and protein metabolism.4,5 It has been found their antioxidant6,7, anti-inflammatory8, antiparasitic9, immunomodulatory10, anabolic4 and cholesterol-lowering7 properties. Another important property of phytoecdysteroids is a reduction in blood glucose level. The blood glucose level had been halved, when alloxan-induced diabetic rats were given complex of phytoecdysteroids isolated from Ajuga iva.7,11 The hypoglycemic property was demonstrated in ecdysone, 2-dexoxyecdysone isolated from the Silene praemixta as well as in 2-deoxyecdisterone and integristeron isolated from Rhaponticum carthamoides.3 Ecdysterone, 22-acetylceiasterone and turkesterone isolated from Ajuga turkestanica also decrease glucose level in intact rats.3 The most expressed hypoglycemic effect among the mentioned above phytoecdysteroids has been found for turkesterone. Its glucose-lowering effect has been well expressed in intact and in adrenaline-, stress- and alloxan-induced hyperglycemia3. In addition to its hypoglycemic property,
turkesterone activates protein synthesis in skeleton muscles, kidneys, and liver. However, there is no information on the effect of turkesterone on the pancreas function in diabetes mellitus.

The study aimed to determine the effect of turkesterone on the endocrine and exocrine parameters of the pancreas in alloxan-induced diabetic rats.

**MATERIALS AND METHODS**

**Animals and experimental design**

Outbred male rats with a weight of 200.0 ± 20.0 g were used in the experiments. Rats were kept in plastic cages of 50x30x28 cm³ in size 4 animals each under natural light and humidity and room temperature. Rats were fed a vivarium diet with unlimited access to water and food. Water and food were changed daily at the same time between 9.00 and 10.00 p.m.

Rats were divided into 4 groups (one control and three experimental). Before modelling alloxan-induced diabetes experimental groups of animals have fasted for two days with unlimited access to water. All experimental group rats were intraperitoneally injected with alloxan monohydrate (DIAEM, OOO, Moscow) solution (170 mg/kg). After alloxan injection, only rats with a serum glucose level of more than 250 mg% were obtained for further experiments. Then the first, second and third experimental groups were intraperitoneally injected with saline, turkesterone (10 mg/kg/24 h) and glibenclamide (5 mg/kg/24 h) (Berlin-Chemie, Germany) respectively for 10 days. Turkesteron (99.5% pure) was isolated from Ajugaturkestanica in the Institute of the Plant Chemistry of the Academy of Sciences of the Republic of Uzbekistan.

Control group animals were given the equivalent volume of saline at the same time and by the same manner. The day when the alloxan-induced diabetic rats began to receive corrective drugs was accepted as the 0th day of observation. Biochemical parameters were determined on the 10th day of observation. For analysis animals were decapitated at 8–10 a.m. Animal procedures were performed according to the Helsinki Declaration of the World Medical Association 2010.

**Preparation of histological and enzymatic active samples**

After decapitation, the pancreas was removed from the abdomen, cleaned from the fat tissue and weighed. Pancreas tail section was cut and stored for 72 h in 10% formalin solution for fixation. Then, the pancreatic section was washed in running water and successively subjected to increased alcohol solutions for dehydration. The preparation was freeze-dried, followed by embedding in paraffin. From each paraffin block, 5–8 μm cross-sections were prepared and stained in hematoxylin-eosin. The preparations were photographed using digital microscopy of the Leake Company (DN-300M). The rest portion of the pancreas was poured with Ringer’s solution (pH 7.4) in 1:9 ratio and homogenized with Teflon pestle at a speed of 400 rpm in a minute. The obtained pancreatic homogenate was centrifuged at 1500 g for 15 min. The supernatant was used to determine the activity of α-amylase (EC 3.2.1.1). All processes were performed in cold conditions.

**Biochemical analysis**

Blood was obtained during rat decapitation into heparin-treated tubes. Blood samples were settled for 30 minutes in a cool place and then centrifuged at 1500 g for 15 minutes. Serum insulin and C-peptide levels were identified with using analyzer Cobas e 411 in the Biochemistry laboratory of the Republican Specialized Scientific-Practical Medical Center of Endocrinology. The blood glucose and total protein level, as well as the activity of α-amylase in the serum and pancreas supernatant, was determined using special reagent sets (Human, Germany).

**Statistical Analysis**

Statistical analyses were conducted using Student t-test. The arithmetic mean (M), standard error (m), t coefficient, and statistical significance value (P) were determined. If the P-value was less than 0.05, the difference between control and experimental animal groups was considered statistically significant.

**RESULTS**

**Pancreas histological structure**

In the control group animals, the pancreatic acinar cells were almost the same size and shape and located linearly. The round shape of endocrine island with numerous cells was occupied a large space. In alloxan-induced diabetic rats exocrine cells vacuolization, enlarging of blood vessels and overgrowth of connective tissue were registered. The endocrine cells were disorganized and some of them were necrotic. However, turkesterone administration notably attenuated the atrophic changes of acinar cells and distraction of endocrine islets. As circulating tissue in the pancreas was improved the endocrine island and acini recovered their morphological state. Administration of glibenclamide to alloxan-induced diabetic rats also resulted in regeneration of the endocrine island and acinar cells (Figure 1).
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**Blood glucose, insulin, C-peptide and total protein level**

Serum glucose level was increased by 3.0 times in alloxan-induced diabetic rats. After daily treatment of alloxan-induced diabetic rats with turkesterone the serum glucose level reached the control level. Treatment with glibenclamide of diabetic rats also resulted in the decrease of glucose level, however, it was statistically higher than the control value.

The insulin level was reduced by almost half in alloxan-induced diabetic rats. Treatment of alloxan-induced diabetic rats with both corrective drugs increased of serum insulin level but did not reach the control value.

A sharp decrease in C-peptide level was observed in alloxan-induced diabetic rats. Treatment of alloxan-induced diabetic rats with turkesterone and/or glibenclamide during 10 days led to partial recovery of serum C-peptide level. Serum total protein level was also decreased in alloxan-induced diabetic rats. When diabetic rats were injected with turkesterone and/or glibenclamide, serum protein level was restored to control value (Table 1).

**Table 1: Effect of turkesterone on the blood glucose, insulin, C-peptide, and total protein level in the alloxan-induced diabetic rat (Mean ± SEM; n=6)**

<table>
<thead>
<tr>
<th>Animal groups*</th>
<th>Glucose (Mmol/l)</th>
<th>Insulin (µE/l)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean ± SEM</td>
<td>%</td>
</tr>
<tr>
<td>C</td>
<td>5.35 ± 0.38</td>
<td>100.00</td>
</tr>
<tr>
<td>AD</td>
<td>15.83 ± 0.86</td>
<td>295.88</td>
</tr>
<tr>
<td>AD+T</td>
<td>6.52 ± 0.45</td>
<td>121.86</td>
</tr>
<tr>
<td>AD+G</td>
<td>6.86 ± 0.58</td>
<td>128.22</td>
</tr>
</tbody>
</table>

**Table 2: Effect of turkesterone on the α–amylase activity in pancreas and serum in alloxan-induced diabetic rats (Mean ± SEM; n = 6)**

<table>
<thead>
<tr>
<th>Animal groups*</th>
<th>Mean ± SEM</th>
<th>Pancreas (U/g tissue)</th>
<th>%</th>
<th>P</th>
<th>Mean ± SEM</th>
<th>Serum (U/l)</th>
<th>%</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>C</td>
<td>36.26 ± 1.42</td>
<td>100.00</td>
<td>-</td>
<td>40.61 ± 2.32</td>
<td>100.00</td>
<td>-</td>
<td></td>
<td></td>
</tr>
<tr>
<td>AD</td>
<td>46.29 ± 2.20</td>
<td>127.66</td>
<td>&lt; 0.002</td>
<td>62.42 ± 2.08</td>
<td>143.86</td>
<td>&lt; 0.01</td>
<td></td>
<td></td>
</tr>
<tr>
<td>AD+T</td>
<td>33.58 ± 2.31</td>
<td>92.61</td>
<td>&gt; 0.25</td>
<td>42.81 ± 1.81</td>
<td>105.42</td>
<td>&gt; 0.5</td>
<td></td>
<td></td>
</tr>
<tr>
<td>AD+G</td>
<td>42.49 ± 2.07</td>
<td>120.54</td>
<td>&lt; 0.02</td>
<td>48.95 ± 2.81</td>
<td>120.54</td>
<td>&lt; 0.05</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>


**α-Amylase activity in pancreas and serum**

In alloxan-induced diabetic rats, pancreas tissue α-amylase activity was significantly increased compared with the non-diabetic rat. The treatment of alloxan-induced diabetic rats with turkesterone resulted in a noticeable decrease in enzyme activity. Administration of glibenclamide also approximated the pancreatic enzyme activity to control level in alloxan-induced diabetic rats. Increase of serum α-amylase activity was clearly expressed in alloxan-induced diabetic rats compared with control. Treatment of alloxan-induced diabetic rats with turkesterone or/ and glibenclamide led to the normalization of serum enzyme activity (Table 2).

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**Figure 1:** Histological structure of rat pancreas (H and E, x400): A. rats treated with saline, B. alloxan-induced diabetic rats, C. alloxan-induced diabetic rats treated with turkesterone, D. alloxan-induced diabetic rats treated with glibenclamide.
DISCUSSION

It is well known that the decrease of insulin and C-peptide levels and the increase of glucose level in serum is the most important diagnostic indicators of diabetes. Such changes take place in patients with type I diabetes and experimental diabetes in animals. Changes of serum parameters accompanied by shifts in the pancreas histostructure.

The observed overgrowth of connective tissue, enlaring of blood vessel volume, swelling of cell and inflammation in pancreas tissue in alloxan-induced diabetic rats can be a consequence of lipid peroxide oxidation caused by increased glucose level. Diabetes depended increase concentration of serum glucose leads to an increase in oxygen-active forms and degradation of lipids, proteins and nucleic acids in cells and organelles. The revealed effect of turkesterone in alloxan-induced diabetic rats probably was due to the anti-inflammatory and antioxidant properties of phytoecdysteroids.

It was shown after treatment of alloxan- or streptozotocin-induced diabetic rats with complex phytoecdysteroids isolated from Ajuga iva the serum glucose and cholesterol levels were reduced, and antioxidant enzyme activity such as catalase, superoxidismutase, glutathioneperoxidase was increased dramatically. Increased activity of antioxidant enzymes under the influence of phytoecdysteroids causes a reduction in lipid peroxide oxidation. It is one of the most important factors in the regulation of insulin secretion and serum glucose level.

Detected in alloxan-induced diabetic rats increasing of the serum α-amylase activity is one of the typical signs of pancreatitis. Increasing the level of pancreas enzymes, including α-amylase, is the main cause of auto-digestion of the gland in pancreatitis. So, autophagy may also play a role in the destruction of pancreatic tissue in alloxan-induced diabetic rats.

The existence of a link between the endocrine and exocrine cell pathologies in the pancreas has been also observed by other authors. Decreased of pancreatic α-amylase releasing in the small intestine, in alloxan-induced diabetic rabbits. This data is associated with our results because in pancreatitis, the releasing of pancreatic enzymes into the intestinal cavity is decreased. Alloxan-induced diabetic rabbits did not recover α-amylase releasing from pancreas cavity in the intestine even after their treatment with insulin. Thus, other non-insulin-dependent changes also occur in the pancreas in alloxan-induced diabetes.

Obtained data show the administration of turkesterone to alloxan-induced diabetic rats cause recovering both endocrine and exocrine function of pancreas. Exocrine pancreas recovery was expressed in the normalization of acinhistostructure and α-amylase activity level in glandular tissue and serum. Wang et al. noted the antiabetic properties of complex phytoecdysteroids isolated from Ajuga iva plant in 28 days after daily treatment of alloxan-induce ddiabetic rats. Obtained data show diabetes correcting effect of turkesterone appears in 10 days. So, it can be suggested the antiabetic effect of turkesterone, isolated from Ajugaturkestanika appears faster than the effect of phytoecdsteroid complex from Ajugaiva. Hence, turkesterone isolated from Ajugaturkestanika has shown the same anti-diabetic effect as glibenclamide. Also, turkestone appears as an anti-pancreatic property. However, the use of turkesterone in clinical practice for the treatment of diabetes and other pancreatic diseases requires additional research.

CONCLUSION

Turkosterone separated from A. Turkestanika has a recovering effect on the endosecretion and exosecretion of the pancreas in alloxan-induced diabetic rats. This is manifested in the restoration of the structure of endocrine islets and exocrine acini, as well as in an increase in insulin and C-peptide levels and a decrease in blood glucose levels in rats with experimental diabetes. Hence, after relevant research, turkesterone can be recommended for the correction of type I diabetes in humans.

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