STUDY OF MICROBIAL FLORA IN PATIENTS WITH INDWELLING CATHETER

Manish N., Tankhiwale N. S.

Department of Microbiology, JNMC Sawangi (M) Wardha (M.S.) India

E-mail of Corresponding Author: nagendra_manish@yahoo.co.in

ABSTRACT

Introduction: - Catheter associated urinary tract infections are common nosocomial infections, 80% cases of UTI are due to use of catheters. The incidence of bacteriuria in catheterized patient is directly related to the duration of catheterization. *E.coli*, *Proteus*, *Pseudomonas*, *Klebsiella*, *Serratia*, *Staphylococcus*, *Enterococci* and *Candida Sp.* Are the common micro-organisms causing this infection? Aims and Objective: - The present study is to determine the microbiological profile and the sensitivity pattern which can cause Catheter Associated Urinary Tract Infections.

Materials and Method: - The study was conducted in 100 adult patients, whom an indwelling Foley’s catheter was inserted in AVBR Hospital during August 2011 to August 2012, in different medical wards, surgery wards and ICU. The catheterized urine sample was collected after catheterization on 5th day onwards on indwelling catheterization. Approximately 3 ml of urine was taken in sterilized container with the sterile precautions. The urine sample microscopy for pus cells, other abnormalities, gram staining was done and inoculate urine in culture medium. The final reading was done after 18 to 24 hrs. Of incubation of culture plates at 37°C. Antibiogram testing was done by Kirby-Bauer disk diffusion technique.

Result and Conclusion: - The most common organism colonizing and causing catheter associated urinary tract infection as per observation were found to be *E.Coli* (57%) followed by *Klebsiella Sp.* (20%), *Staphylococcus* (8%), *Enterococcus Sp.* (6%), *Pseudomonas aeruginosa* (5%) and *Acinetobacter Sp.* (4%). Frequent cleanliness of catheter, area to avoid contamination and colonization of microbial flora, it is recommended to change the catheter every 5th day.

Keywords: - Catheter associated urinary tract infection, bacteriuria, *E.Coli*

INTRODUCTION

Catheter associated urinary tract infections are common nosocomial infections, 80% cases of UTI are due to use of catheters.¹ Fifteen to twenty five percent of patient in hospitals, need catheterization.² Factors that have increased the risk of catheter associated urinary tract infections are prolonged catheterization, severe underlying illness, disconnection of catheter and drainage tube, faulty catheter care and lack of systemic antibiotics therapy.³ About 1% to 48% of hospitalized patient with indwelling catheters still acquire the infection.² The incidence of bacteriuria in catheterized patient is directly related to the duration of catheterization.⁴ Urethral catheter is a major predisposing factor in the development of nosocomial UTI and catheter- associated bacteremia.⁵ Most nosocomial UTI can be benign but a systemic complication which is gram-negative septicemia can develop in 30-40% of patients.

*E.coli*, *Proteus*, *Pseudomonas*, *Klebsiella*, *Serratia*, *Staphylococcus*, *Enterococci* and *Candida Sp.* are the common micro-organisms causing this infection. Many infecting strains display markedly greater antibiotics resistance.
than organisms that cause community-acquired Urinary tract infection. Study of microbial flora in these patients can prevent establishment of UTI and further kidney damage. Antibiotic sensitivity testing will help the clinician to give proper treatment.

**AIMS AND OBJECTIVE**

The present study is undertaken to determine the microbiological profile and the sensitivity pattern of the strains which can cause Catheter Associated Urinary Tract Infections.

**MATERIAL AND METHOD**

The study was conducted in 100 adult patients, whom an indwelling Foley’s catheter was inserted in AVBR Hospital during August 2011 to August 2012. The study was undertaken in patients catheterized and admitted in different medical wards, surgery wards and ICU. The catheterized urine sample was collected after catheterization on 5th day onwards on indwelling catheterization. Sample was collected by 24-gauge needle with all sterile precautions and urine was brought to microbiology lab within 1 hour of collection for further processing. Approximately 3 ml of urine was taken as a sample in sterilized container with the sterile precautions with sterilized syringe. The micro-organism growth was seen only after 5th day of catheterization.

The urine sample microscopy for pus cells, other abnormalities and gram staining was done. Standard loop was used for inoculating urine in culture medium. The urine was subjected to culture on Blood agar, MacConkey agar, CLED (Cystine Lactose Electrolyte Deficient Medium), TSI and Nutrient agar. The final reading was done after 18 to 24 hrs. of incubation of culture plates at 37°C. The identification of micro-organisms was done by the colony characters, Gram staining, morphology, biochemical reactions as per standard text book (Practical Medical Microbiology, Mackie & McCartney, 14th edition).

Colony count $>10^5$ cfu/ml was considered as significant. Antibiogram testing was done by Kirby-Bauer disk diffusion technique in Mueller-Hinton medium.

**DISCUSSION**

The result of the microbiologic profile in this study is similar to most reported studies. *E.Coli* still being the most common pathogen (57% of cases) followed by *Klebsiella pneumoniae*(20%), *Staphylococcus*(8%), *Enterococcus Sp.*(6%), *Pseudomonas aeruginosa *(5%) and *Acinetobacter Sp.*(4%).

In a study conducted by Poudel C.M., Baniya G. Department of Internal Medicine and Department of Microbiology, TUTH, Katmandu the majority of organisms belonged to *E.Coli* (40.77%), *Klebsiella pneumoniae* (11.11%), *Enterococcus Sp.*(11.11%), *Pseudomonas* (11.11%), *Acinetobacter Sp.* (3.7%) are all most similar. Present study is comparable to those workers.

The most common organism colonizing and causing catheter associated urinary tract infection as per observation were found to be *E.Coli* (57%) followed by *Klebsiella Sp.*(20%), *Staphylococcus* (8%), *Enterococcus Sp.* (6%), *Pseudomonas aeruginosa* (5%) and *Acinetobacter Sp.* (4%).

*E.Coli* are found to be sensitive to Amikacin (87.71%), Nitrofurantoin (82.46%) Ceftazidime (14.03%), least sensitive to Ciprofloxacin (8.77%) and Co-trimaxazole (7.02%) and 100% sensitive to Imipenem.

*Klebsiella Sp.* are found to be sensitive to Amikacin (75%), Ciprofloxacin (30%), least sensitive to Nitrofurantoin (9%) and 100% sensitive to Imipenem and 100% resistance to Co-trimoxazole.

*Staphylococcus* are found to be sensitive to Ciprofloxacin (75%), Co-trimoxazole (62.5%), Erythromycin (62.5%) and 100% sensitive to...
Nitrofurantoin and Vancomycin and 100% resistance to Penicillin.

*Enterococcus Sp.* are found to be sensitive to Nitrofurantoin (66.66%), Amikacin (33.33%) and 100% sensitive to Vancomycin and Lenezolid and 100% resistance to Penicillin and Ciprofloxacin.

*Pseudomonas aeruginosa* are found to be sensitive to Amikacin (40%), Netillin (40%) and 100% sensitive to Imipenam and Pipracillin and 100% resistance to Ceftazidime, Nitrofurantoin and Ciprofloxacin.

Acinetobacter Sp. are found to be sensitive to Amikacin (50%) and 100% sensitive to Imipenam and 100% resistance to Ceftazidime, Nitrofurantoin, Ciprofloxacin and Co-trimoxazole.

**CONCLUSION AND RECOMMENDATIONS**

- Frequent cleanliness of catheter, area to avoid contamination and colonization of microbial flora, it is recommended to change the catheter every 5th day.
- Proper aseptic precaution to be taken while collection of samples by syringe.
- Microbial flora should be evaluated in catheterized patients through out.
- The antibiotic sensitivity report should be referred before giving treatment, if infection is established.

**ACKNOWLEDGEMENT**

We acknowledge the great help received from the scholars whose articles cited and included in references of this manuscript. We are also grateful to authors / editors / publishers of all those articles, journals and books from where the literature for this article has been reviewed and discussed. We are grateful to IJCRR editorial board members and IJCRR team of reviewers who have helped to bring quality to this manuscript.

**REFERENCES**

OBSERVATIONS

Table No. 1, Micro organism grown in catheterized urine samples (n=100) culture Positive after 5 days of catheterization.

<table>
<thead>
<tr>
<th>S.No</th>
<th>Bacteria Grown in Cultures</th>
<th>No. of samples (n=100)</th>
<th>Percentages(%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.</td>
<td><em>E. Coli</em></td>
<td>57</td>
<td>57</td>
</tr>
<tr>
<td>2.</td>
<td><em>Klebsiella pneumoniae</em></td>
<td>20</td>
<td>20</td>
</tr>
<tr>
<td>3.</td>
<td><em>Staphylococcus</em></td>
<td>08</td>
<td>8</td>
</tr>
<tr>
<td>4.</td>
<td><em>Enterococci Sp.</em></td>
<td>06</td>
<td>6</td>
</tr>
<tr>
<td>5.</td>
<td><em>Pseudomonas aeruginosa</em></td>
<td>05</td>
<td>5</td>
</tr>
<tr>
<td>6.</td>
<td><em>Acinetobacter Sp.</em></td>
<td>04</td>
<td>4</td>
</tr>
</tbody>
</table>

Table No. 2 – Antibiotic Sensitivity pattern of Isolated Organisms in Percentage (%)

<table>
<thead>
<tr>
<th>Organisms</th>
<th>Antibiotics</th>
<th>Imipenem</th>
<th>Amikacin</th>
<th>Cefazidime</th>
<th>Nitrofurantoin</th>
<th>Ciprofloxacinecin</th>
<th>Co-trimoxazole</th>
<th>Netilimiprin</th>
<th>Pipracillin</th>
<th>Vancomycin</th>
<th>Lenezolid</th>
<th>Erythromycin</th>
<th>Penicillin</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>E. coli</em></td>
<td></td>
<td>100</td>
<td>87.71</td>
<td>14.03</td>
<td>82.46</td>
<td>8.77</td>
<td>7.02</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Klebsiella pneumoniae</em></td>
<td></td>
<td>100</td>
<td>75</td>
<td>20</td>
<td>9</td>
<td>30</td>
<td>0</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Staphylococcus</em></td>
<td></td>
<td>100</td>
<td>75</td>
<td>62.5</td>
<td></td>
<td>100</td>
<td>62.5</td>
<td>0</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Enterococci sp.</em></td>
<td></td>
<td>33.33</td>
<td>66.66</td>
<td>0</td>
<td>100</td>
<td>100</td>
<td>0</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Pseudomonas aeruginosa</em></td>
<td></td>
<td>100</td>
<td>40</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>40</td>
<td>100</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Acinetobacter Sp.</em></td>
<td></td>
<td>100</td>
<td>50</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>